Tap water embedding and ultra-rapid methylene blue stain for fast Mohs micrographic surgery processing

R. Moro B. Gallardo-Sanz M.J. Roca-Estellés L. Alfaro-Ferreres

PII: S0001-7310(25)00096-1

DOI: https://doi.org/doi:10.1016/j.ad.2024.09.029

Reference: AD 4266

To appear in: Actas dermosifiliograficas

Received Date: 2 September 2024
Accepted Date: 22 September 2024

Please cite this article as: Moro R, Gallardo-Sanz B, Roca-Estellés MJ, Alfaro-Ferreres L, Tap water embedding and ultra-rapid methylene blue stain for fast Mohs micrographic surgery processing, *Actas dermosifiliograficas* (2025), doi: https://doi.org/10.1016/j.ad.2024.09.029

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2025 Published by Elsevier España, S.L.U. on behalf of AEDV.



Sección: Vídeo en Cirugía Dermatológica

Tap water embedding and ultra-rapid methylene blue stain for fast Mohs micrographic

surgery processing

Incrustación en agua del grifo y tinción ultrarrápida con azul de metileno para un

procesamiento veloz de la cirugía micrográfica de Mohs

Authors:

R. Moro^{1,2}; B. Gallardo-Sanz³; M. J. Roca-Estellés³; and L. Alfaro-Ferreres³

Institutions:

¹Escuela de Doctorado, Universidad Católica de Valencia San Vicente Mártir, València, Spain

²Instituto Dermatológico Tekderma, Hospital Vithas Valencia 9 de Octubre, València, Spain;

³Servicio de Patología, Hospital Vithas Valencia 9 de Octubre, València, Spain

Corresponding author:

Ruggero Moro,

E-mail address: ruggero_moro@hotmail.com

Mohs micrographic surgery (MMS) is known to be a time- and resource-consuming procedure; therefore, methods to optimize MMS workflow are always welcome.

In this brief communication we wanted to share our way to perform MMS processing using tap water (TW) embedding and methylene blue (M-blue) stain. As far as we know, both TW embedding and M-blue stain have never been reported for MMS processing. However, they have been historically used for the intraoperative pathologic evaluation of other surgical specimens(1, 2). Therefore, the application of this processing method to MMS is a "back to basics" of intraoperative pathology. Feasibility of this processing is supported by the data of 261 consecutive MMS cases collected in a prospective database from February 2022 through August 2024.

Standard optimal cutting temperature (OCT) embedding involves using embedding devices, embedding molds, or glass slides and a spirit lever or a heat extractor. The reason to use OCT is to quickly reach an optimal cutting temperature to avoid well-known frozen artifacts. However, in our experience TW does not generate more frozen artifacts than OCT with faster freezing times.

Our MMS processing follows these steps: 1) after surgical excisions, MMS specimens are processed as per standard MMS technique; 2) a cryostat tissue holder (chuck) is prepared in the cryostat with a layer of frozen TW (ie, an ice bed) to have a smooth surface to attach and lay the sections flat; 3) the chuck is taken out from the cryostat and the MMS section is attached to it; 4) drops of TW are added around the section to complete the embedding; 5) the chuck is, then, put back in the cryostat and the heat extractor is applied; 6) after 10–15 seconds, the section is ready to be trimmed and cut. Trimming is usually reduced because there is no layer of OCT or other embedding medium to remove. The Mohs technician places progressive layers on 1 glass slide making sure he gets the entire section. Finally, slides are stained with M-blue with the following ultra-rapid protocol: 5 seconds immersion in M-blue (Kühne's Methylene Blue Phenicated solution, QCA, Amposta, ES) and 5 seconds wash in TW. Slides are, then, dried with paper and passed to the pathologist and Mohs surgeon to be evaluated. To further speed up the processing, no cover slip is applied, and no mounting media is needed. They will be applied later when the slides are ready to be scanned (Aperio GT450, Leica, Wetzlar, DE). The steps of this MMS processing and some tips performance-wise are shown video 1. Quality of the slides obtained with our method is shown in Fig. 1-2 and Video 1.

The use of TW embedding can be a reason form concern in some readers. Such concern also worried the main author (R.M.) when he started performing MMS after his fellowship. However, after more than 2 years, we agree that using TW embedding does not negatively affect the quality of the slides. Indeed, it seems to be very helpful for cutting through adipose tissue, a known problem

of MMS(3). Therefore, TW embedding maintains frozen section skin quality, while enhancing fat quality without having to further freez with ice spray or liquid nitrogen [Fig. 1-2]. Moreover, it reduces trimming time because no layer of embedding medium covers the specimen surface. Finally, using TW instead of OCT lowers the costs of MMS processing.

One drawback of TW embedding may be the difficulty to keep the MMS section flattened once flipped over on the chuck, because it will not be "floating" in the OCT. Indeed, the skin edge may drop below the horizontal plane of the section, especially if cutting a single-section MMS. Nevertheless, the skin edges can be elevated [Video 1]. Comparison between TW and OCT embedding is shown in Table 1.

The use of M-blue stain may be less surprising. M-blue stain is practically identical to toluidine blue (T-blue) stain. T-blue is very well known in literature for enhancing visualization of skin cancers normally treated by MMS. This enhancement is due to a phenomenon called metachromasia, whereby the T-blue staining of the mucopolysaccharide stroma around tumor cells shifts towards pink or magenta(4). Although M-blue shows the same metachromatic phenomenon as T-blue [Fig. 1 and Video 1], it has never been reported as a stain of MMS layers. There are not clear advantages or drawbacks associated with the use M-blue instead of T-blue; it is just a matter of preference, cost, and availability. Remarkably, our 10 seconds M-blue protocol is way faster than previously described T-blue protocols(4, 5).

Although there have not been recurrences of skin cancers treated with this MMS processing, follow-up time—2.5 years—is very short to draw any conclusions on recurrence rates.

In conclusion, we reported how we do the MMS processing illustrating 2 novel features: TW embedding and ultra-rapid M-blue stain. These new features speed up MMS processing without seemingly affecting slides quality and clinical results. Review of the slides without mounting media or cover slips further speeds up the process, which is especially important when multiple sections are required. Longer follow-up and comparative studies are needed to confirm our experience.

Ética de la publicación

1. ¿Su trabajo ha comportado experimentación en animales?:

No

2. ¿En su trabajo intervienen pacientes o sujetos humanos?:

No

3. ¿Su trabajo incluye un ensayo clínico?:

No

4. ¿Todos los datos mostrados en las figuras y tablas incluidas en el manuscrito se recogen en el apartado de resultados y las conclusiones?:

Sí

References:

- 1. WILSON LB. A METHOD FOR THE RAPID PREPARATION OF FRESH TISSUES FOR THE MICROSCOPE. Journal of the American Medical Association 1905;XLV:1737-37.
- 2. Arteta JL. [Urgent biopsy; Rapid tissue diagnosis during operative intervention]. Rev Esp Cir (Madr 1944) 1945;1:209-18.
- 3. Reserva J, Kozel Z, Krol C, et al. Processing Adipose-Rich Mohs Samples: A Comparative Study of Effectiveness of Pretreatment With Liquid Nitrogen Versus Flash Freezing Spray. Am J Dermatopathol 2017;39:838-41.
- 4. Todd MM, Lee JW, Marks VJ. Rapid toluidine blue stain for Mohs' micrographic surgery. Dermatol Surg 2005;31:244-5.
- 5. Donaldson MR, Weber LA. Toluidine Blue Supports Differentiation of Folliculocentric Basaloid Proliferation From Basal Cell Carcinoma on Frozen Sections in a Small Single-Practice Cohort. Dermatol Surg 2017;43:1303-06.

Figure legends:

Figure 1. Metachromasia is visible as a violet halo around 2 foci (263 nm/pixel, upper 7x, lower 10x) of a nodular basal cell carcinoma in this final layer of a 2-section Mohs micrographic surgery specimen (263 nm/pixel, 1.08x). Slide quality is good and adipose tissue is present.

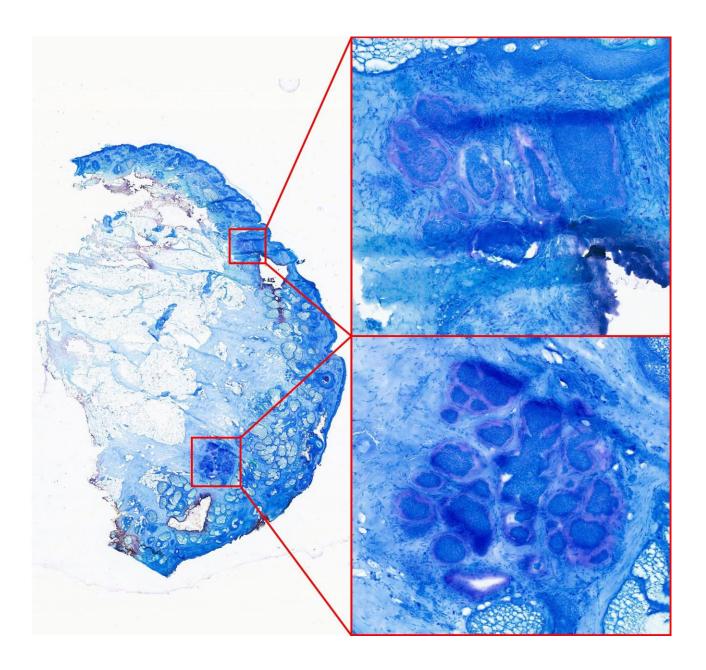


Figure 2. Slide quality of a large (3 cm) adipose rich Mohs micrographic surgery specimen (263 nm/pixel, 0.52x). At 5x the excellent frozen fat quality is evident.

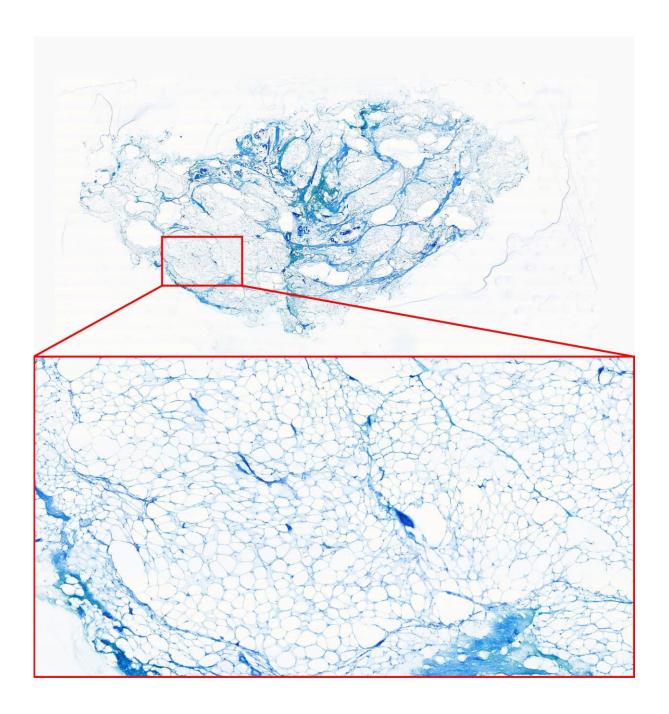


Table 1. Comparison between tap water (TW) and optimal cutting temperature (OCT) embedding

	TW embedding	OCT embedding
	• Faster freezing time (10-15	Standard embedding medium
	seconds)	
Pros	Good frozen fat quality without	
	further freezing	
	No cost	
	Embedding and cutting may be	• Slower freezing time (> 60 seconds)
Cons	more technically challenging	Frozen fat quality may be low
		without further freezing