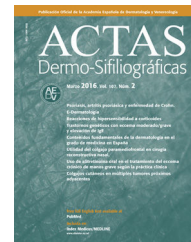




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REVIEW

Epidermal Nevi and Related Syndromes — Part 1: Keratinocytic Nevi[☆]



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PALABRAS CLAVE

Nevus epidérmico;
Nevus
queratinocítico;
*Papular Epidermal
Nevus with Skyline
basal cell layer*
(PENS)

Abstract Epidermal nevi are hamartomatous lesions derived from the epidermis and/or adnexal structures of the skin; they have traditionally been classified according to their morphology. New variants have been described in recent years and advances in genetics have contributed to better characterization of these lesions and an improved understanding of their relationship with certain extracutaneous manifestations. In the first part of this review article, we will look at nevi derived specifically from the epidermis and associated syndromes.

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Nevus epidérmicos y síndromes relacionados. Parte 1: nevus queratinocíticos

Resumen Los nevus epidérmicos son hamartomas originados en la epidermis y/o en las estructuras anexiales de la piel que se han clasificado clásicamente partiendo de la morfología. En los últimos años se han descrito variantes nuevas y se han producido avances en el campo de la genética que han permitido caracterizar mejor estas lesiones y comprender su relación con algunas de las manifestaciones extracutáneas a las que se han asociado. En esta primera parte revisaremos los nevus derivados de la epidermis y los síndromes que se han descrito asociados a ellos.

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Introduction

A hamartoma is a tumor-like malformation, usually congenital, arising due to an abnormal mixture in the distribution or proportions of mature, constitutive tissue elements. It is not a neoplasm because the tissues do not undergo autonomous growth. Skin hamartomas are called nevi. There is a

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Table 1 Epidermal Nevi Classified According to Morphological Criteria and Genes Implicated in Development.

		Gene involved	Locus		
Keratinocytic nevi	Common keratinocytic nevus	FGFR3	4p16.3		
		PIK3CA	3q26.32		
		HRAS	11p15.5		
		NRAS	1p13.2		
		KRAS	12p12.1		
		FGFR2	10q26.13		
		Unknown	Unknown		
		PENS	Unknown		
		Epidermal nevus in Proteus syndrome	AKT1	14q32.33	
		Type 2 segmental Cowden disease (PTEN)	PTEN	10q23.31	
		CHILD nevus	NSDHL	Xq28	
		ILVEN	Unknown	Unknown	
		Epidermolytic epidermal nevus	KRT1	12.q13.13	
	KRT10	17q21.2			
	ATPA2A	12q24.11			
Nevi derived from adnexal structures	Nevus sebaceous	HRAS	11p15.5		
		NRAS	1p13.2		
		KRAS	12p12.1		
	Follicular nevi	Hair follicle nevus (congenital vellus hamartoma)	FGFR2 ^a	10q26.13	
			Unknown	Unknown	
			NEK9, FGFR2	14q24.3, 10q26.13	
			Unknown	Unknown	
	Apocrine nevi	Apocrine nevus	Unknown	Unknown	
			Syringocystadenoma papilliferum	BRAF	7q34
			Eccrine nevi	Eccrine nevus	Unknown
	Eccrine nevi	Eccrine angiomas hamartoma	Unknown	Unknown	
			Porokeratotic adnexal ostial nevus	GJB2	13q12.11
			Becker nevus	ACTB	7p22.1

Abbreviations: HILD, congenital hemidysplasia, ichthyosiform erythroderma and limb defects; ILVEN, inflammatory linear verrucous epidermal nevus; PENS, papular epidermal nevus with skyline basal cell layer; PTEN, in this context, papillomatous, thick, epidermal, nonorganoid nevus.

^a Association of mutations of the *FGFR2* gene with sebaceous nevus has been debated (see text).

certain degree of confusion regarding the definition of the term *epidermal nevus*. Most articles in the scientific literature consider epidermal nevi to refer to lesions derived from the epidermis or from adnexal epithelial cells. Some texts on skin pathology, however, define epidermal nevi as those lesions derived from epidermal keratinocytes (herein denoted keratinocytic nevi) excluding nevi derived from adnexal structures.^{1,2} In this article, we will refer to both nevi derived from the epidermis and those derived from adnexal structures. The review will be divided into 2 parts. **Table 1** summarizes and classifies the nevi that will be reviewed both in part 1 (keratinocytic nevi) and in part 2 (nevi derived from adnexal structures) and can be used as a table of contents.

Epidermal nevi often exhibit so-called organocity. This concept, in pathology, defines the simultaneous growth of several cell components in the same hamartoma. This phenomenon is often observed in hamartomas derived from adnexal structures and should not be confused with the concept, more often used in medicine, to define a

symptom originating from organic and physical abnormality (as opposed to a psychological cause). Some texts classify epidermal nevi as organoid and nonorganoid (in practice, keratinocytic). Adnexal structures, for their part, can be the cause of certain lesions defined in some skin pathology text books as hamartomas,³ but which are, however, not considered nevi in most text books on clinical dermatology (for example, steatocystoma, fibrofolliculoma, and trichofolliculoma). We will not cover these lesions in the present review.

Pathophysiological Basis

The appearance of these lesions is due to genetic mutations or epigenetic changes that impact the expression of a cell clone during embryo development, leading to mosaicism, that is, the presence of 2 or more genetically distinct cell populations in the same individual.⁴ Four genetic mechanisms have been described that explain most cases of mosaicism in the skin:

- 1) Somatic mutations in autosomal (dominant) genes that are lethal when they occur in the zygote but persist in the mosaic form. These mutations affect only one group of cells, which survive because they reside close to normal cells.⁵ Most epidermal nevi occur due to mutations of this type in 2 interlinked signaling pathways implicated in survival and cell proliferation: the phosphatidylinositol 3 kinase (PI3K) pathway and the mitogen-activated protein kinase or MAP-kinase pathway.⁶
- 2) Nonlethal mutations in autosomal genes that cause extensive skin disease when they occur in the zygote, but that can also become manifest in mosaic form if they are the result of a postzygotic mutation. These mutations can occur in an otherwise healthy individual, comprising an exclusively mosaic involvement (type 1 Happle mosaicism) or in a patient who already presents a generalized form of the disease, with a greater area of involvement, generally due to an additional mutation that leads to loss of heterozygosity (type 2 mosaicism).
- 3) Mosaic mutations or epigenetic changes in genes linked to polygenic inflammatory diseases.
- 4) Functional mosaicism linked to random inactivation of 1 of the X chromosomes in women, also known as lyonization.

Table 1 lists the genes whose mutation has been implicated in the pathogenesis of epidermal nevi. Recent discoveries in the field of genetics have shown that linking genotype to phenotype is far more complex than expected.⁷ The appearance of keratinocytic nevi may be due to mutations in up to 6 different genes, without any relationship with specific morphological findings. On the other hand, the same mutation in the same gene may give rise to morphologically distinct lesions, as occurs with mutations in RAS genes. Such mutations have been described in the pathogenesis of both keratinocytic and sebaceous nevi, even in the same patient.

Skin hamartomas follow a distribution pattern that depends on the type of cell from which they are derived.^{8,9} Most epidermal nevi follow the Blaschko lines (Happle pattern A and B archetype) with certain exceptions. Furthermore, the timing of the mutation and the onset of mosaicism determines the site and extent of the nevus. When the onset of mosaicism occurs towards the end of development, the lesions are limited and adopt different shapes⁹: an oval or triangular form is most frequent in epidermal nevi. The most extensive lesions result from mosaicism that occurred earlier and are more often associated with extracutaneous involvement. The epidermal nevus syndrome or Solomon syndrome was a term coined for certain patients in whom an

Table 2 Some of the Best Characterized Syndromes Associated With Keratinocytic Nevi.

Syndrome	Type of Nevus	Other Manifestations	Gene Involved (Transmission)	Reference
Epidermal nevus syndrome FGFR3 (García-Happle)	Keratinocytic nevus	Mental retardation, epilepsy	<i>FGFR3</i> (sporadic)	29,30
CLOVES syndrome	Keratinocytic nevus	Lipomatous overgrowth, vascular, skeletal abnormalities	<i>PIK3CA</i> (sporadic)	32
PENS syndrome	Keratinocytic nevus (PENS)	Mild mental and psychomotor retardation that tends to improve with age, Achilles tendon shortening, hypospadias, curved penis	Unknown (most are sporadic, some familial cases with paradominant inheritance)	46–48
Proteus syndrome	Keratinocytic nevus	Limb hypertrophy, macrodactyly, vascular malformations, lipomas, cutis aplasia congenita and connective tissue nevus, lung disease, parotid adenoma, ovarian cystadenoma and breast cancer, endometrial cancer, testicular cancer	<i>AKT1</i> (sporadic)	50,51
Type 2 segmental Cowden disease	Keratinocytic nevus (PENS)	Trichilemmomas, oral papillomas, fibromas, mucocutaneous neuromas, acral keratosis, genital lentiginosis, malformations, and vascular tumors and lipomas. Breast cancer, endometrial cancer, thyroid cancer, and colon cancer	<i>PTEN</i> (autosomal dominant)	53,55,56
CHILD syndrome	Keratinocytic nevus (CHILD nevus)	Ipsilateral skeletal aplasia or hypoplasia, punctiform calcifications in the bone ends, ipsilateral brain, lung, heart, or kidney defects	<i>NSDHL</i> (dominant X-linked, although the majority are sporadic)	58

Abbreviations: CHILD, congenital hemidysplasia, ichthyosiform erythroderma and limb defects; CLOVES, congenital lipomatous overgrowth with vascular, epidermal and skeletal anomalies; PENS, papular epidermal nevus with skyline basal cell layer; PTEN, in this context, papillomatous, thick, epidermal, nonorganoid nevus.



Figure 1 Linear epidermal nevus following the Blaschko lines.

epidermal nevus was associated with defects in the central nervous system, eyes, and/or bones.¹⁰ This terminology does not adequately describe the different entities that have been discovered in recent years and that are genetically and clinically very heterogeneous.¹¹⁻¹⁴ Table 2 summarizes the most important keratinocytic nevus syndromes described to date.

Common Keratinocytic Nevus

The common keratinocytic nevus presents as brown or grey plaques with a verrucous or velvety surface, often distributed along the Blaschko lines (Fig. 1). Their prevalence is between 0.1% and 0.5%. The lesions located in skinfolds can become moist and malodorous, whereas those on extensor surfaces often become dry and cracked. Some lesions may have a notably papillomatous appearance (Fig. 2). We often refer to these as verrucous nevi. The term nevus unius lateris refers to long linear keratinocytic nevi, generally on a limb. These are uncommon on the scalp and forehead, where sebaceous nevi are much more frequent. Some epidermal



Figure 2 Verrucous nevus. Some keratinocytic nevi acquire a clearly papillomatous surface.

nevi occur in association with woolly hair on the scalp.¹⁵ Histologically, common keratinocytic nevi present acanthosis, papillomatosis, hyperkeratosis, thickening of the granular layer, and increased melanin in the basal layer (Fig. 3). Some histopathological variants have been described.^{16,17}

The appearance of malignant tumors on keratinocytic nevi is uncommon, although cases have been published (Fig. 4).¹⁸⁻²⁴ They usually occur in adults. The most common is the development of squamous cell carcinoma. In addition, cases have been reported of basal cell carcinoma and porocarcinoma.

Common keratinocytic nevi are the result of mutations in up to 6 different genes, and so these may not translate into specific morphological features. All these are protooncogenes implicated in some types of cancer as well as in other benign acquired lesions such as seborrheic keratosis.²⁵ We will now review the syndromes associated with keratinocytic nevi, based on their genetic classification.

Genetic Classification of Keratinocytic Nevi and Associated Syndromes

In approximately one third of common keratinocytic nevi, the same R248C heterozygous mutation in the *FGFR3* oncogene can be found.^{25,26} *FGFR3* mutations are responsible for a relatively long list of syndromes and also neoplasms such as urinary bladder cancer, uterine cervical cancer, colorectal cancer, and spermatocytic seminoma. Although many cases of urinary tract cancer have been reported in patients with keratinocytic nevi,^{27,28} there is no proof to date that the nevus and neoplasm share the same mutation in any patients. In 2 patients with epidermal nevus and urothelial cancer in whom the *FGFR3* gene was studied, both the hamartoma and neoplasm were wild type.²⁹ Mosaic mutations in *FGFR3* have been reported in some patients with mental retardation and epilepsy. When this occurs, the condition is known as the *FGFR3* syndrome or García-Hafner-Happle syndrome.^{30,31}

Another gene often affected is the *PIK3CA* gene. The same E545G mosaic mutation of *PIK3CA* has been implicated in keratinocytic nevi, seborrheic keratosis, and colorectal cancer.³² *PIK3CA* mutations have also been associated with lipomatous growth and vascular and skeletal abnormalities, grouped in a syndrome known as congenital lipomatous overgrowth with vascular, epidermal and skeletal anomalies (CLOVES), Online Mendelian Inheritance in Man (OMIM) #612918.³³ This syndrome was described in a series of 7 patients previously diagnosed with Proteus syndrome who had certain differential characteristics: progressive and mixed complex vascular malformations on the trunk, adipose tissue abnormalities, variable degrees of scoliosis, and elongated bones without progressive overgrowth and without bone distortion characteristic of Proteus syndrome.³⁴ Subsequently, patients were described with malformations of the central nervous system and epilepsy.³⁵ Unlike patients with Proteus syndrome, they did not develop connective tissue nevus (elastoma, collagenoma).³⁶ Linear keratinocytic nevus is, in contrast, a characteristic lesion in CLOVES syndrome. The lesions tend to grow and become more verrucous until adolescence, and are stable thereafter.

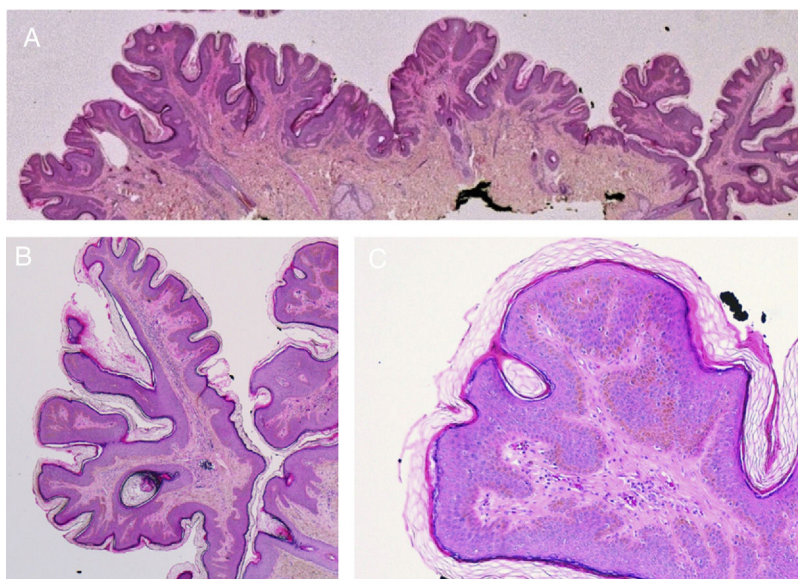


Figure 3 Verrucous nevus on the back of the neck. Histological image. A, Hematoxylin-eosin (HE) 2x: the epidermis shows acanthosis, papillomatosis, and hyperkeratosis. B, HE, 4x: detail of the papillomatous epidermis. C, HE, 10x: greater detail of the acanthosis, presence of hyperpigmentation of the basal layer, and *loosely woven basket* orthokeratotic hyperkeratosis.

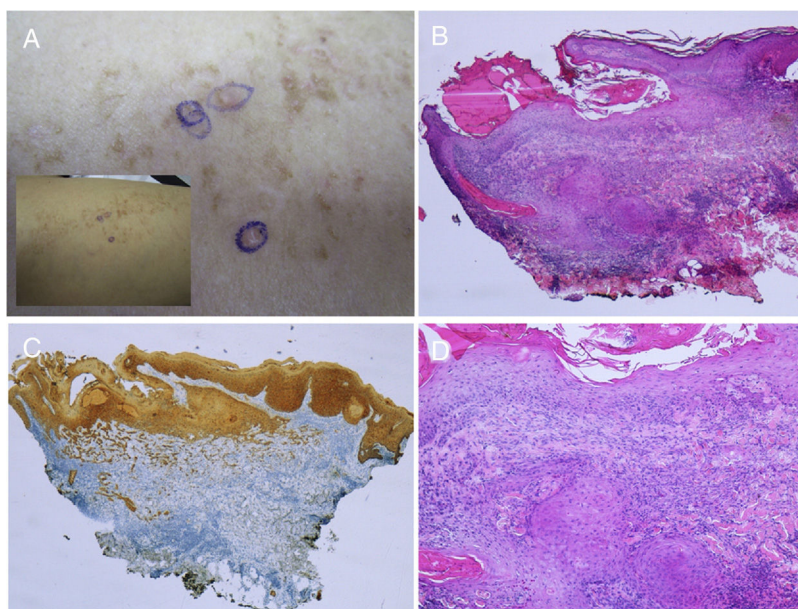


Figure 4 Squamous cell carcinoma developed over epidermic nevus, previously treated with electrocoagulation. A, Low resolution and detailed photograph. B, Hematoxylin-eosin (HE), 4x: 2 of the 3 lesions that were biopsied showed an epidermal proliferation and formation of strands that invaded the reticular dermis. C, Immunohistochemistry with AE1/AE3 cytokeratin, 4x: these strands of epithelial cells were invading the dermis. D, HE, 10x detail of the neoplastic proliferation composed of atypical keratinocytes that form keratin and horny pearls. Clinical images with permission of Dr. M. Concepción Sánchez Bermejo and Dr. María José González, of the Hospital de Manacor.

Approximately 40% of keratinocytic nevi are the result of RAS mutations. In a study of 72 lesions, the *HRAS* gene was the most frequently affected, followed by *NRAS* and *KRAS*.³⁷ RAS genes are important oncogenes and carry mutations in up to 30% of neoplasms in humans and in multiple syndromes (RASopathies). In the group of epidermal nevi, mosaic mutations in this group of genes may give rise to both keratinocytic nevi and organoid nevi and are an example of discordance between genotype and phenotype. Nevus

marginatus, which has a central area rich in sebaceous glands and a more papular peripheral area with acanthosis and papillomatosis more similar to a keratinocytic nevus, is caused by mutations in RAS genes present in both components.³⁸ Recently, the case has been reported of a girl with multiple nevoid lesions, some of which were consistent with keratinocytic nevus on the trunk and others with nevus sebaceous in the craniofacial region.³⁹ The patient also showed delayed tooth development, cerebral arachnoid

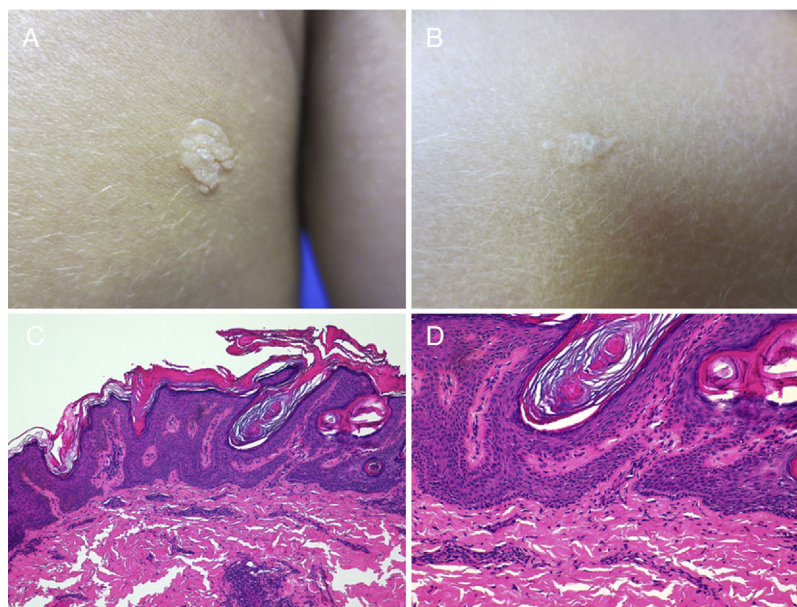


Figure 5 Two PENS in the same patient. A and B, Two plaques, of 8 mm and 5 mm, with rounded form in the first case and polygonal somewhat linear form in the second. C, Hematoxylin-eosin (HE), 4 x: biopsy showed epidermal hyperplasia with thickened and merged crests, with papillomatosis and compact hyperkeratosis. D, HE, 10x: at higher magnification, the basal cell layer in the epidermis can be seen with a cobblestone appearance.

cyst, and optical atrophy. In both types of nevus, the same *KRAS* mutation was found. This mutation was not present in healthy skin or in peripheral blood of her parents. The syndromic forms associated with mutations in RAS genes will be covered in greater detail in the review of sebaceous nevus.

Some keratinocytic nevi with RAS mutations have been associated with the development of malignant neoplasms that carry the same mutation. This occurred for example in the case of a girl who presented a keratinocytic nevus and developed uterovaginal rhabdomyosarcoma at the age of 6 months.⁴⁰ Both the hamartoma and the neoplasm showed the same *KRAS* mutation, which was not present in other tissues of the same patient. Another case is a 49-year-old man with extensive keratinocytic nevus who developed urothelial cancer in which the same mosaic *HRAS* mutation was detected in both lesions.⁴¹

Very recently, abnormalities have been described in the *FGFR2* oncogene, whose mutation would explain 5% to 10% of keratinocytic nevi.⁴² The recent description of 2 fetuses with sebaceous nevi that carry mutations of this gene has been the source of debate, as some authors consider that these are keratinocytic nevi.^{43,44}

Other Keratinocytic Epidermal Nevi

Papular Epidermal Nevus with Skyline Basal Cell Layer (PENS) and PENS Syndrome

In 2011, Antonio Torrelo et al.⁴⁵ described a new form of epidermal nevus with specific clinical and histological features, corresponding to solitary or multiple, congenital lesions or lesions with onset soon after birth, with a papular appearance and smooth or papillomatous surface, forming plaques that are rounded or with form of a comma, measuring 0.3

to 1.5 cm with a random distribution. Only one case has been reported in which the lesions followed a Blaschkoid distribution.⁴⁶ Histologically, they present regular epidermal acanthosis with thickened and rectangular epidermal crests, with orthokeratotic hyperkeratosis. The most characteristic finding is that the basal cell layer is arranged in a cobblestone pattern, with an empty eosinophilic region between the nuclei of the basal layer and the first keratinocytes of the stratum spinosum (Fig. 5).

PENS syndrome is manifest in the form of mental retardation and mild delay in development of psychomotor skills and epilepsy in the first year of life.⁴⁷ These manifestations usually improve or even remit as the child gets older. Other findings include characteristic facies, shortened Achilles heel tendon, hypospadias, and curved penis.^{48,49}

The genetic mutation responsible for this disorder and the mode of transmission are not known. In the original description, *FGFR3* and *PIK3CA* mutations were ruled out.⁴⁵ Generally, the lesions are sporadic, although families have been reported with more than one member affected.^{47,50}

Epidermal Nevus in Proteus Syndrome

Proteus syndrome (OMIM #176920) is characterized by asymmetric tissue overgrowth caused by lethal, nonhereditary, mutations of the *AKT1* gene that survive in mosaic form.⁵¹ The most characteristic changes are macrodactyly and hypertrophy of the limbs. Vascular malformations, lipomas, aplasia cutis congenita and connective tissue nevus, lung disease, parotid adenoma, ovarian cystadenoma and breast cancer, endometrial cancer, and testicular cancer may also be present. Up to 50% of patients present with epidermal nevi. These are flattened solitary or multiple plaques with a

velvety surface, following the Blaschko lines. Histologically, acanthosis and hyperkeratosis are observed. Unlike other tumors associated with Proteus syndrome, epidermal nevus does not grow over time.⁵²

Before the discovery of *AKT1* mutations, Proteus-like syndrome due to mutations in the *PTEN* gene, with autosomal dominant transmission, has led to some confusion. Currently, it is thought that all these patients have Cowden disease along with an epidermal nevus (see the next section).^{53,54} All cases of Proteus syndrome are sporadic.

Type 2 Segmental Cowden Disease (PTEN nevus)

Cowden syndrome (OMIM #158350) is a multisystemic disorder characterized by the appearance of multiple hamartomas and tumors, including breast cancer, endometrial cancer, thyroid cancer, and colon cancer. Tumors of the skin include trichilemmomas, oral papillomas, fibromas (including sclerotic fibroma or storiform collagenoma, which seems to be quite a specific finding), mucocutaneous neuromas, acral keratosis, genital lentiginosis, malformations, and vascular tumors and lipomas. It is produced by a mutation in the *PTEN* gene, a tumor suppressor gene of autosomal dominant transmission, although mutations have been reported in other genes.⁵⁵

The lesion has also been called Cowden nevus or PTEN (used here as the acronym for papillomatous, thick, epidermal, nonorganoid nevus), and refers to the appearance of erythematous, hyperkeratotic and papillomatous papules that coalesce to form linear plaques following the Blaschko lines in patients with Cowden syndrome. It is caused by loss of heterozygosity in the *PTEN* gene (type 2 Happle mosaicism).^{54,56} It has also been called type 2 segmental Cowden disease,⁵⁴ segmental overgrowth, lipomatosis, arteriovenous malformation and epidermal nevus syndrome.⁵⁷ The lesion can be distinguished from the epidermal nevus characteristic of Proteus syndrome because of its more keratotic and papillomatous appearance and because it presents with other manifestations of Cowden syndrome.

CHILD Nevus and CHILD Syndrome

Congenital hemidysplasia, ichthyosiform erythroderma, and limb defects (CHILD) syndrome (OMIM #308050) is characterized by an epidermal nevus in association with ipsilateral skeletal aplasia or hypoplasia, punctiform calcifications in bone ends, ipsilateral defects in the brain, lungs, heart, or kidney. It is transmitted according to an X-linked dominant pattern although most cases are sporadic. Clinical expression is very variable, even within the same family, and members may have very subtle lesions that hinder diagnosis.⁵⁸ The disease arises as a result of mutation of the *NSDHL* gene,⁵⁹ which participates in cholesterol metabolism. The mutation is very often lethal in male embryos, although cases have been described in males.⁶⁰

CHILD nevus is often confused with an inflammatory disease, particularly psoriasis, in cases with more limited involvement.⁶¹ It presents as linear erythematous plaques or yellowish, scaling, psoriasiform or ichthyosiform plaques, following a Blaschkoid distribution. The most extensive

lesions have a well-defined border at the midline. Interestingly, the great majority of patients described have involvement of the right half of the body. There is a particular predilection for folds (ptychotropism).⁶² Histologically, the disease is characterized by psoriasiform dermatitis with neutrophil exocytosis. In parakeratosis, it has been noted that the nuclei that persist are more rounded compared with those that are seen in psoriasis.⁶¹ We often see accumulations of histiocytes carrying lipid vacuoles that express CD68 and adipophilin in the papillary dermis, which under a papillomatous epidermis, may lead to suspicion of a verruciform xanthoma. These findings, in the context of a congenital Blaschkoid, lateral lesion, with ptychotropism, should lead to suspicion of CHILD syndrome.

Inflammatory Linear Verrucous Epidermal Nevus (ILVEN)

This lesion is a linear plaque with an inflammatory appearance and erythema and scaling. It tends to present on the legs. In 25% of cases, the lesions are present at birth, a further 50% present during the first 6 months, and the remaining lesions can present up until the patient is 4 years of age. Histologically, we see a psoriasiform dermatitis, with epidermal hyperplasia and areas of parakeratosis on an epidermis without a granular layer, alternating with orthokeratosis over hypergranulosis. Areas with a more spongiotic appearance with lymphocyte exocytosis have been described. The disease can present in the form of type 1 or type 2 mosaicism in patients with generalized psoriasis.⁶³ Several cases have been reported with good response to treatments for psoriasis (topical agents, phototherapy, systemic agents, biologics). Apart from an anecdotic association with arthropathy,⁶⁴ extracutaneous manifestations have not been described in patients with linear epidermal verrucous inflammatory nevus. The genetic mutations responsible for the mosaicism giving rise to this nevus are not known, and, like psoriasis, it could be a polygenic disease.

Epidermolytic Keratinocytic Nevus

This disease corresponds to a mosaic form of epidermolytic ichthyosis or epidermolytic hyperkeratosis (OMIM #113800). It is due to dominant mutations in keratin 1 or 10 (*KRT 1* or *KRT10*).^{65,66} These are somewhat pigmented verrucous lesions following a linear Blaschkoid distribution. They may be present at birth as a solitary lesion or multiple lesions, on any part of the body. The flexural lesions often become moist and malodorous. Histologically, they present hyperkeratosis, acanthosis, papillomatosis, and acantholysis in the granular layer. These findings are consistent with epidermolytic ichthyosis. Cases of type 2 mosaicism have been reported, with linear areas of greater involvement on an individual with this disease.⁶⁷ Several families have been reported with parents affected with the nevoid form and children who present a complete form of epidermolytic ichthyosis.⁶⁸⁻⁷⁰ It is suspected that transmission occurs through gonadal mosaicism. In patients in whom this nevus is diagnosed, germinal involvement should be ruled out and genetic counselling made available.

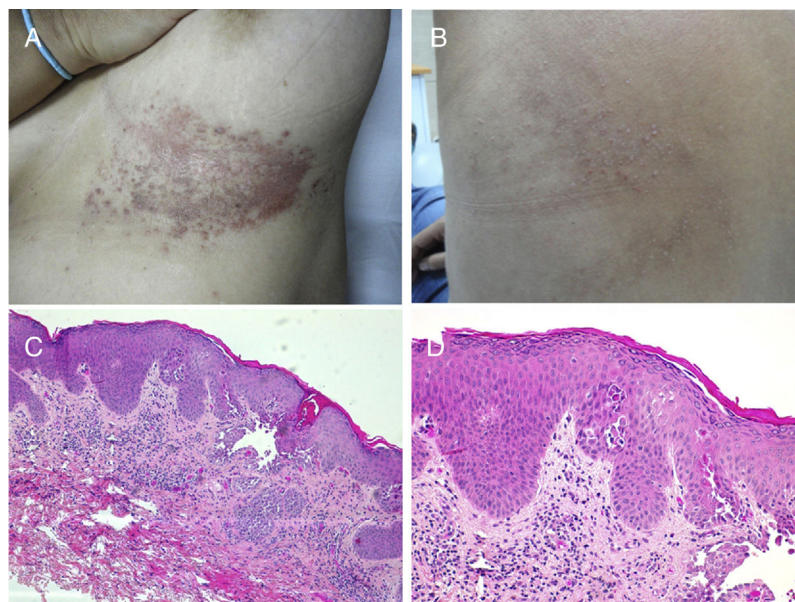


Figure 6 Segmental Darier disease. A and B, Clinical images of the submammary and left dorsal lesion in a female patient without any additional skin lesions. C, Hematoxylin-eosin (HE), 4x: biopsy showed epidermal acanthosis with minimal orthokeratotic hyperkeratosis, suprabasal acantholysis, and formation of an intraepidermal blister. D, HE 10x: at higher magnification, the dyskeratotic changes can be appreciated better, with presence of rounded bodies and spots. The patient was diagnosed with segmental Darier disease.

Acantholytic and Dyskeratotic Epidermal Nevus

As in the previous nevus, most authors consider this nevus a mosaic form of a monogenic disease, although there is certain disagreement on this point. In this case, it would correspond to nevoid forms of Darier disease (OMIM #124200) or more rarely Hailey-Hailey disease (OMIM #169600). Numerous cases of segmental or linear Darier disease have been reported in the literature such as segmental or linear Darier disease.⁷¹⁻⁷⁵ Clinically, these are linear keratotic crusty lesions that appear at puberty or later (Fig. 6). Exacerbations are characteristic and often associated with sweating and exposure to sunlight. Mucosal manifestations and lesions on the limbs, including palms, soles, and nails, characteristic of Darier disease, are not usually seen in nevoid forms. Furthermore, the lack of a family history of Darier disease is characteristic.

Histologically, we see the presence of a hyperkeratotic and parakeratotic epidermis in which we observe intraepidermal cleavage that contains acantholytic and dyskeratotic keratinocytes, with presence of corps ronds and grains (Fig. 6). In some cases of acantholytic and dyskeratotic epidermal nevus, a mutation has been found in *ATP2A2*, characteristic of the disease.^{76,77}

Some authors affirm, however, that acantholytic and dyskeratotic epidermal nevus and segmental Darier disease are not the same entity.⁷⁸⁻⁸⁰ The examples presented are congenital lesions (instead of lesions developing after puberty), in which mutation in the *ATP2A2* gene has not been demonstrated or it is possible to confirm expression of SERCA2 (the expression product of *ATP2A2*) by immunohistochemistry.

Nevoid forms of Hailey-Hailey disease are rare. One case has been published of type 1 segmental involvement,⁸¹

and another case with presentation at 3 months of age with a relapsing linear lesion that subsequently developed symmetric lesions characteristic of the disease.⁸² In this patient, loss of heterozygosity was demonstrated in the area affected earliest (type 2 mosaicism).⁸³ Histologically, suprabasal acantholysis is observed with lymphocytes and eosinophils. Dyskeratosis can be present, although it is not usually as evident as in Darier disease and rounded bodies or grains are not usually observed. Diagnosis of Hailey-Hailey disease usually requires immunofluorescence to rule out an autoimmune blistering disease, particularly, pemphigus vulgaris.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

1. Patterson J. *Weedon's Skin Pathology*. 4th edition Edinburgh: Elsevier; 2015.
2. Calonje E, Brenn T, Lazar A, McKee PH. *McKee's Pathology of the Skin*. 4th edition Philadelphia: Editorial Saunders Elsevier; 2012.
3. Requena L. *Neoplasias anexas cutáneas*. Madrid: Aula Médica Ediciones; 2004.
4. Happle R. What is a nevus? A proposed definition of a common medical term. *Dermatology*. 1995;191:1-5.
5. Rodríguez Bandera AI, Feito Rodríguez M, Vorlinka K, de Lucas Laguna R. Líneas de Blaschko y otros mosaicismos cutáneos. *Piel*. 2013;28:457-68.
6. Asch S, Sugarman JL. Epidermal nevus syndromes: New insights into whorls and swirls. *Pediatr Dermatol*. 2018;35:21-9.

7. Has C. Mosaicism in the skin: lumping or splitting? *Br J Dermatol.* 2017;176:15–6.
8. Happle R. Mosaicism in human Skin: understanding the patterns and mechanisms. *Arch Dermatol.* 1993;129:1460–70.
9. Torrelo A, Baselga E, Nagore E, Zambrano A, Happle R. Delineation of the various shapes and patterns of nevi. *Eur J Dermatol.* 2005;15:439–50.
10. Solomon LM, Fretzin DF, Dewald RL. The epidermal nevus syndrome. *Arch Dermatol.* 1968;97:273–85.
11. Happle R. How many epidermal nevus syndromes exist? A clinicogenetic classification. *J Am Acad Dermatol.* 1991;25:550–6.
12. Happle R. Epidermal nevus syndromes. *Semin Dermatol.* 1995;14:111–21.
13. Happle R. The group of epidermal nevus syndromes Part I Well defined phenotypes. *J Am Acad Dermatol.* 2010;63:1–22.
14. Happle R. The group of epidermal nevus syndromes Part II Less well defined phenotypes. *J Am Acad Dermatol.* 2010;63:25–30.
15. Martín-González T, del Boz-González J, Vera-Casaño A. Nevus de pelo lanoso asociado a nevus epidérmico lineal ipsilateral. *Actas Dermosifilogr.* 2007;98:198–201.
16. Su WPD. Histopathologic varieties of epidermal nevus. A study of 160 cases. *Am J Dermatopathol.* 1982;4:161–70.
17. Submoke S, Piamphongsant T. Clinico-histopathological study of epidermal naevi. *Australas J Dermatol.* 1983;24:130–6.
18. Dogliotti M, Frenkel A. Malignant change in a verrucous nevus. *Int J Dermatol.* 1978;17:225–7.
19. Cramer SF, Mandel MA, Hauler R, Lever WF, Jenson AB. Squamous cell carcinoma arising in a linear epidermal nevus. *Arch Dermatol.* 1981;117:222–4.
20. Horn MS, Sausker WF, Pierson DL. Basal cell epithelioma arising in a linear epidermal nevus. *Arch Dermatol.* 1981;117:247.
21. Levin A, Amazon K, Rywlin AM. A squamous cell carcinoma that developed in an epidermal nevus. *Am J Dermatopathol.* 1984;6:51–5.
22. Ichikawa T, Saiki M, Kaneko M, Saida T. Squamous cell carcinoma arising in a verrucous epidermal nevus. *Dermatology.* 1996;193:135–8.
23. Affleck AG, Leach IH, Varma S. Two squamous cell carcinomas arising in a linear epidermal naevus in a 28-year-old female. *Clin Exp Dermatol.* 2005;30:382–4.
24. Masood Q, Narayan D. Squamous cell carcinoma in a linear epidermal nevus. *J Plast Reconstr Aesthet Surg.* 2009;62:693–4.
25. Hafner C, Toll A, Fernández-Casado A, Earl J, Marqués M, Acquadro F, et al. Multiple oncogenic mutations and clonal relationship in spatially distinct benign human epidermal tumors. *Proc Natl Acad Sci U S A.* 2010;107:20780–5.
26. Hafner C, van Oers JM, Vogt T, Landthaler M, Stoehr R, Blaszyk H, et al. Mosaicism of activating FGFR3 mutations in human skin causes epidermal nevi. *J Clin Invest.* 2006;116:2201–7.
27. Rongioletti F, Rebora A. Epidermal nevus with transitional cell carcinomas of the urinary tract. *J Am Acad Dermatol.* 1991;25:856–8.
28. Rosenthal D, Fretzin DF. Epidermal nevus syndrome: report of association with transitional cell carcinoma of the bladder. *Pediatr Dermatol.* 1986;3:455–8.
29. Hernández S, Toll A, Baselga E, Ribé A, Azua-Romeo J, Pujol RM, et al. Fibroblast growth factor receptor 3 mutations in epidermal nevi and associated low grade bladder tumors. *Invest Dermatol.* 2007;127:1664–6.
30. García-Vargas A, Hafner C, Pérez-Rodríguez AG, Rodríguez-Rojas LX, González-Esqueda P, Stoehr R, et al. An epidermal nevus syndrome with cerebral involvement caused by a mosaic FGFR3 mutation. *Am J Med Genet A.* 2008;146A:2275–9.
31. Ousager LB, Bygum A, Hafner C. Identification of a novel S249C FGFR3 mutation in a keratinocytic epidermal nevus syndrome. *Br J Dermatol.* 2012;167:202–4.
32. Hafner C, López-Knowles E, Luis NM, Toll A, Baselga E, Fernández-Casado A, et al. Oncogenic PIK3CA. mutations occur in epidermal nevi seborrheic keratoses with a characteristic mutation pattern. *Proc Natl Acad Sci U.S.A.* 2007;104:13450–4.
33. Kurek KC, Luks VL, Ayturk UM, Alomari AI, Fishman SJ, Spencer SA, et al. Somatic mosaic activating mutations in PIK3CA cause CLOVES syndrome. *Am J Hum Genet.* 2012;90:1108–15.
34. Sapp JC, Turner JT, van de Kamp JM, van Dijk FS, Lowry RB, Biesecker LG. Newly delineated syndrome of congenital lipomatous overgrowth, vascular malformations, and epidermal nevi (CLOVE syndrome) in seven patients. *Am J Med Genet A.* 2007;143:2944–58.
35. Gucev ZS, Tasic V, Jancevska A, Konstantinova MK, Pop-Jordanova N, Trajkovski Z, et al. Congenital lipomatous overgrowth, vascular malformations, and epidermal nevi (CLOVE) syndrome: CNS malformations and seizures may be a component of this disorder. *Am J Med Genet A.* 2008;146A:2688–90.
36. Martínez-Lopez A, Blasco-Morente G, Perez-Lopez I, Herrera-García JD, Luque-Valenzuela M, Sanchez-Cano D, et al. CLOVES syndrome: review of a PIK3CA-related overgrowth spectrum (PROS). *Clin Genet.* 2017;91:14–21.
37. Hafner C, Toll A, Gantner S, Mauerer A, Lurkin I, Acquadro F, et al. Keratinocytic epidermal nevi are associated with mosaic RAS mutations. *J Med Genet.* 2012;49:249–53.
38. Groesser L, Vogt T, Happle R, Herschberger E, Mickler M, Landthaler M, et al. Naevus marginatus revisited: a combined organoid and nonorganoid epidermal naevus caused by HRAS mutation. *Br J Dermatol.* 2013;168:892–4.
39. Igawa S, Honma M, Minami-Hori M, Tsuchida E, Iizuka H, Ishida-Yamamoto A. Novel postzygotic KRAS mutation in a Japanese case of epidermal nevus syndrome presenting with two distinct clinical features, keratinocytic epidermal nevi and sebaceous nevi. *J Dermatol.* 2016;43:103–4.
40. Bourdeaut F, Héroult A, Gentien D, Pierron G, Ballet S, Reynaud S, et al. Mosaicism for oncogenic G12D KRAS mutation associated with epidermal nevus, polycystic kidneys and rhabdomyosarcoma. *J Med Genet.* 2010;47:859–62.
41. Hafner C, Toll A, Real FX. HRAS mutation mosaicism causing urothelial cancer and epidermal nevus. *N Engl J Med.* 2011;365:1940–2.
42. Toll A, Fernández LC, Pons T, Groesser L, Sagrera A, Carrillo-de Santa Pau E, et al. Somatic Embryonic FGFR2 Mutations in Keratinocytic Epidermal Nevi. *J Invest Dermatol.* 2016;136:1718–21.
43. Kuentz P, Fraitag S, Gonzales M, Dhombres F, St-Onge J, Duffourd Y, et al. Mosaic-activating FGFR2 mutation in two fetuses with papillomatous pedunculated sebaceous naevus. *Br J Dermatol.* 2017;176:204–8.
44. Asch S, Sugarman JL. 52 words for snow: Dermatologists naming epidermal nevi. *Br J Dermatol.* 2018;178:296.
45. Torrelo A, Colmenero I, Kristal L, Navarro L, Hafner C, Hernández-Martín A, et al. Papular epidermal nevus with «skyline» basal cell layer (PENS). *J Am Acad Dermatol.* 2011;64:888–92.
46. Faure E, Tadini G, Brena M, Cassulini LR. Papular epidermal nevus with «skyline» basal cell layer (PENS) following a Blaschko linear pattern. *Pediatr Dermatol.* 2013;30:e270–1.
47. Tadini G, Restano L, Happle R, Itin P. PENS syndrome: a new neurocutaneous phenotype. *Dermatology.* 2012;224:24–30.
48. Rodríguez-Díaz E, Gonzalvo P, Colmenero I, Requena L, Hernández-Martín A, Torrelo A. Papular epidermal nevus with «skyline» basal cell layer (PENS) with extracutaneous findings. *Pediatr Dermatol.* 2013;30:e54–6.
49. Luna PC, Pannizardi AA, Martín CI, Vigovich F, Casas JG, Larrale M. Papular Epidermal Nevus with Skyline Basal Cell Layer (PENS): three new cases and review of the literature. *Pediatr Dermatol.* 2016;33:296–300.

50. Brena M, Besagni F, Boneschi V, Tadini G. Familial papular epidermal nevus with «skyline» basal cell layer. *Pediatr Dermatol.* 2014;31:e33–5.
51. Lindhurst MJ, Sapp JC, Teer JK, Johnston JJ, Finn EM, Peters K, et al. A mosaic activating mutation in AKT1 associated with the Proteus syndrome. *N Engl J Med.* 2011;365:611–9.
52. Twede JV, Turner JT, Biesecker LG, Darling TN. Evolution of skin lesions in Proteus syndrome. *J Am Acad Dermatol.* 2005;52:834–8.
53. Cohen MM Jr, Turner JT, Biesecker LG. Proteus syndrome: misdiagnosis with PTEN mutations. *Am J Med Genet A.* 2003;122A:323–4.
54. Happle R. Type 2 segmental Cowden disease vs Proteus syndrome. *Br J Dermatol.* 2007;156:1089–90.
55. Ni Y, Zbuk KM, Sadler T, Patocs A, Lobo G, Edelman E, et al. Germline mutations and variants in the succinate dehydrogenase genes in Cowden and Cowden-like syndromes. *Am J Hum Genet.* 2008;83:261–8.
56. Happle R. Linear Cowden nevus: a new distinct epidermal nevus. *Eur J Dermatol.* 2007;17:133–6.
57. Caux F, Plauchy H, Chibon F, Faivre L, Fain O, Vabres P, et al. Segmental overgrowth, lipomatosis, arteriovenous malformation and epidermal nevus (SOLAMEN) syndrome is related to mosaic PTEN nullizygoty. *Eur J Hum Genet.* 2007;15:767–73.
58. Bittar M, Happle R, Grzeschik KH, Leveleki L, Hertl M, Bornholdt D, et al. CHILD syndrome in 3 generations: the importance of mild or minimal skin lesions. *Arch Dermatol.* 2006;142:348–51.
59. König A, Happle R, Bornholdt D, Engel H, Grzeschik KH. Mutations in the NSDHL gene, encoding a 3beta-hydroxysteroid dehydrogenase, cause CHILD syndrome. *Am J Med Genet.* 2000;90:339–46.
60. Happle R, Effendy I, Megahed M, Orlow SJ, Küster W. CHILD syndrome in a boy. *Am J Med Genet.* 1996;62:192–4.
61. Gantner S, Rütten A, Requena L, Gassenmaier G, Landthaler M, Hafner C. CHILD syndrome with mild skin lesions: histopathologic clues for the diagnosis. *J Cutan Pathol.* 2014;41:787–90.
62. Happle R. Ptychotropism as a cutaneous feature of the CHILD syndrome. *J Am Acad Dermatol.* 1990;23:763.
63. Hofer T. Does inflammatory linear verrucous epidermal nevus represent a segmental type 1/type 2 mosaic of psoriasis? *Dermatology.* 2006;212:103–7.
64. Al-Enezi S, Huber AM, Krafchik BR, Laxer RM. Inflammatory linear verrucous epidermal nevus and arthritis: a new association. *J Pediatr.* 2001;138:602–4.
65. Tsubota A, Akiyama M, Sakai K, Goto M, Nomura Y, Ando S, et al. Keratin 1 gene mutation detected in epidermal nevus with epidermolytic hyperkeratosis. *J Invest Dermatol.* 2007;127:1371–4.
66. Samuelov L, Sarig O, Gat A, Halachmi S, Shalev S, Sprecher E. Extensive lentigo simplex, linear epidermolytic naevus and epidermolytic naevus comedonicus caused by a somatic mutation in KRT10. *Br J Dermatol.* 2015;173:293–6.
67. Eng AM, Brody P, Rhee HL, Bronson DM. Congenital ichthyosiform erythroderma and epidermal nevus. *Int J Dermatol.* 1991;30:284–7.
68. Nazzaro V, Ermacora E, Santucci B, Caputo R. Epidermolytic hyperkeratosis: generalized form in children from parents with systematized linear form. *Br J Dermatol.* 1990;122:417–22.
69. Paller AS, Syder AJ, Chan YM, Yu QC, Hutton E, Tadini G, et al. Genetic and clinical mosaicism in a type of epidermal nevus. *N Engl J Med.* 1994;331:1408–15.
70. Chassaing N, Kanitakis J, Sportich S, Cordier-Alex MP, Titeux M, Calvas P, et al. Generalized epidermolytic hyperkeratosis in two unrelated children from parents with localized linear form, and prenatal diagnosis. *J Invest Dermatol.* 2006;126:2715–7.
71. Chester BJ, Brown L. Darier's disease resembling linear verrucous epidermal nevus. *Arch Dermatol.* 1959;80:625–6.
72. Cambiaghi S, Brusasco A, Grimalt R, Caputo R. Acantholytic dyskeratotic epidermal nevus as a mosaic form of Darier's disease. *J Am Acad Dermatol.* 1995;32:284–6.
73. Gilaberte M, Puig L, Vidal D, Alomar A. Acantholytic dyskeratotic naevi following Blaschko's lines: a mosaic form of Darier's disease. *J Eur Acad Dermatol Venereol.* 2003;17:196–9.
74. Martínez S, Vera A, Eloy-García C, Sanz A, Crespo V. Enfermedad de Darier lineal. *Actas Dermosifiliogr.* 2006;97:139–41.
75. De la Hera I, Chico R, Llamas R, Vanaclocha F. Enfermedad de Darier segmentaria. *Actas Dermosifiliogr.* 2011;102:299–301.
76. Sakuntabhai A, Dhitavat J, Burge S, Hovnanian A. Mosaicism for ATP2A2 mutations causes segmental Darier's disease. *J Invest Dermatol.* 2000;115:1144–7.
77. Wada T, Shirakata Y, Takahashi H, Murakami S, Iizuka H, Suzuki H, et al. A Japanese case of segmental Darier's disease caused by mosaicism for the ATP2A2 mutation. *Br J Dermatol.* 2003;149:185–8.
78. Mazereeuw-Hautier J, Thibaut I, Bonafé JL. Acantholytic dyskeratotic epidermal nevus: a rare histopathologic feature. *J Cutan Pathol.* 2002;29:52–4.
79. Huh WK, Fujiwara K, Takahashi H, Kanitakis J. Congenital acantholytic dyskeratotic epidermal naevus following Blaschko's lines versus segmental Darier's disease. *Eur J Dermatol.* 2007;17:130–2.
80. Akinshemoyin Vaughn O, Hinshaw MA, Teng JM. Acantholytic dyskeratotic epidermal nevus. *JAMA Dermatol.* 2015;151:1259–60.
81. Hwang LY, Lee JB, Richard G, Uitto JJ, Hsu S. Type 1 segmental manifestation of Hailey-Hailey disease. *J Am Acad Dermatol.* 2003;49:712–4.
82. Vakilzadeh F, Kolde G. Relapsing linear acantholytic dermatosis. *Br J Dermatol.* 1985;112:349–55.
83. Poblete-Gutiérrez P, Wiederholt T, König A, Jugert FK, Marquardt Y, Rübber A, et al. Allelic loss underlies type 2 segmental Hailey-Hailey disease, providing molecular confirmation of a novel genetic concept. *J Clin Invest.* 2004;114:1467–74.