



# ACTAS Dermo-Sifiliográficas

Full English text available at  
[www.actasdermo.org](http://www.actasdermo.org)



## REVIEW

# Cutaneous and Mucocutaneous Leishmaniasis<sup>☆</sup>

I. Abadías-Granado,<sup>\*</sup> A. Diago, P.A. Cerro, A.M. Palma-Ruiz, Y. Gilaberte



Servicio de Dermatología, Hospital Universitario Miguel Servet, Zaragoza, Spain

Received 29 December 2020; accepted 13 February 2021  
Available online 11 May 2021

### KEYWORDS

Cutaneous leishmaniasis;  
Mucocutaneous leishmaniasis;  
Clinical manifestations;  
Diagnosis;  
Treatment

**Abstract** Leishmaniasis is a chronic disease caused by flagellate protozoa of the genus *Leishmania*. It is a global disease, but most cases are seen in South America, the Mediterranean, and some areas of Asia and Africa. The 3 main types of leishmaniasis are cutaneous (the most common), mucocutaneous, and visceral (the most severe). Visceral leishmaniasis is also known as *kala-azar*. Leishmaniasis is diagnosed by demonstrating the presence of *Leishmania* amastigotes in clinical specimens using direct microscopic examination or molecular analysis. Various treatments exist, although the evidence supporting the options available for cutaneous leishmaniasis is weak. Both the classical presentation of leishmaniasis and our management of the disease have changed in recent decades because of acquired immune deficiency caused by conditions such as human immunodeficiency infection or the use of tumor necrosis factor inhibitors.  
© 2021 Published by Elsevier España, S.L.U. on behalf of AEDV. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

### PALABRAS CLAVE

Leishmaniasis cutánea;  
Leishmaniasis mucocutánea;  
Manifestaciones clínicas;  
Diagnóstico;  
Tratamiento

### Leishmaniasis cutánea y mucocutánea

**Resumen** La leishmaniasis es una enfermedad crónica causada por un protozoo flagelado perteneciente al género *Leishmania*. Tiene distribución mundial, aunque la mayoría de casos se agrupan en América del Sur, la cuenca mediterránea y algunas zonas de Asia y África. Existen tres formas fundamentales de enfermedad: cutánea, la más frecuente; mucocutánea; y visceral, también denominada *kala-azar*, la forma más grave. El diagnóstico se establece con la demostración de la presencia de los amastigotes en muestras clínicas, mediante visión directa al microscopio o mediante técnicas moleculares. Existen múltiples opciones terapéuticas, aunque la evidencia en la que se basa el tratamiento de la leishmaniasis cutánea es débil. Actualmente, las alteraciones de la inmunidad producidas por factores como el VIH o el uso de fármacos anti-TNF han cambiado tanto la forma de presentación de las formas clínicas clásicas como sus tratamientos.  
© 2021 Publicado por Elsevier España, S.L.U. en nombre de AEDV. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

<sup>☆</sup> Please cite this article as: Abadías-Granado I, Diago A, Cerro PA Palma-Ruiz AM, Gilaberte Y. Leishmaniasis cutánea y mucocutánea. ACTAS Dermo-Sifiliogr. 2021;112:601–618.

<sup>\*</sup> Corresponding author.

E-mail addresses: [isabel.abadiasg@gmail.com](mailto:isabel.abadiasg@gmail.com), [iabadias@salud.aragon.es](mailto:iabadias@salud.aragon.es) (I. Abadías-Granado).

## Introduction

Leishmaniasis is a chronic disease caused by flagellate protozoa of the genus *Leishmania*, of which there are over 20 species. *Leishmania* parasites are obligate intracellular parasites transmitted by the bite of infected female sandflies of the *Phlebotomus* and *Lutzomyia* genera. Leishmaniasis is essentially a zoonotic disease. The main reservoirs are dogs and rodents, but there are 2 cases for which humans are the main reservoirs: *Leishmania donovani* and *Leishmania tropica*.

## Epidemiology

The estimated incidence of leishmaniasis according to the World Health Organization is 700 000 to 1 million cases a year, of which 50 000 to 90 000 correspond to visceral leishmaniasis (VL).<sup>1</sup> Approximately 95% of all cutaneous leishmaniasis (CL) cases occur in South America, the Mediterranean Basin, the Middle East, or Central Asia. LV, by contrast, is predominant in Brazil, East Africa, and India. In 2018, over 85% of new CL cases reported to the WHO were from Afghanistan, Algeria, Bolivia, Brazil, Colombia, Iran, Iraq, Pakistan, Syria, and Tunisia, while over 95% of new LV cases were from Brazil, China, Ethiopia, India, Iraq, Kenya, Nepal, Somalia, and Sudan.<sup>1</sup> Finally, over 90% of new mucocutaneous leishmaniasis (MCL) cases were reported in 4 countries: Brazil, Bolivia, Ethiopia, and Peru.<sup>1</sup>

Leishmaniasis has traditionally been classified as Old World or New World depending on its geographic location. Old World leishmaniasis occurs in Asia, Africa, and Europe, and is mostly caused by *L tropica*, *Leishmania major*, *Leishmania aethiopica*, *Leishmania infantum*, or *L donovani*. New World leishmaniasis, in turn, occurs in America, and is mostly caused by *Leishmania mexicana*, *Leishmania amazonensis*, *Leishmania braziliensis*, *Leishmania panamensis*, or *Leishmania infantum chagasi* (a subspecies of *L infantum* in the New World, formerly known as *Leishmania chagasi*).<sup>2</sup> The main epidemiological and clinical patterns for most common *Leishmania* species are shown in Table 1.<sup>2-5</sup> Fig. 1 shows the geographic distribution of the main species and their principal reservoirs.

In Spain, leishmaniasis is a zoonotic disease endemic to the peninsula and the Balearic Islands. The main causative species in both CL and VL is *L infantum*, which is transmitted by *Phlebotomus perniciosus* and *Phlebotomus ariasi*.<sup>6</sup> Dogs are the main reservoir for *L infantum*, although other reservoirs such as hares<sup>7</sup> and rats<sup>8</sup> have been described. Apart from its endemic form, leishmaniasis can occur in patients with human immunodeficiency virus (HIV) infection<sup>9</sup> or a compromised immune system (e.g., patients being treated with tumor necrosis factor [TNF] inhibitors).<sup>10</sup> There have also been epidemic outbreaks, such as the Madrid outbreak that occurred between 2009 and 2013.<sup>7</sup> Finally, phenomena such as globalization, international travel, and migration have increased the prevalence of leishmaniasis in developed countries.<sup>11</sup>

## Pathogenesis

*Leishmania* promastigotes are injected into human skin by infected female sandflies. Here, through phagocytosis by macrophages, they transform into amastigotes, multiplying inside the cells and infecting other mononuclear phagocytized cells. Sandflies become infected by ingesting infected cells while feeding on the host's blood. Once in the intestine of the sandflies, the amastigotes transform into promastigotes.<sup>12</sup> The incubation period of leishmaniasis varies according to the clinical form of disease, but is generally 2 weeks (or less) to 2 months for CL, 3 to 9 months for VL, and over 2 years for MCL.<sup>13</sup>

The clinical manifestations of leishmaniasis vary according to the species involved<sup>14</sup> (Table 1) and the host's immune response.<sup>15</sup> The spectrum of immune response ranges from a strong T-cell response that results in the production of interferon (IFN)  $\gamma$  to a humoral response that produces high antibody levels. *Leishmania* species are eliminated by IFN- $\gamma$ -activated macrophages, but cannot be neutralized by antibodies. This is why individuals who mount a strong immune response have lesions with few parasites, while those with a humoral response are unable to control infection. Diffuse CL is caused by uncontrolled infection. It should also be noted that an exaggerated T helper type 1 response and increased expression of CD8<sup>+</sup> T cells are associated with more severe forms of disease, such as MCL.<sup>15</sup>

## Clinical Manifestations

### Cutaneous Leishmaniasis

CL starts with the formation of a papule at the inoculation site, which is typically located in exposed areas of the body, such as the face or extremities. The papule typically develops into a plaque or a nodule (Fig. 2A-E) with a tendency to ulcerate. Gum collectors (*chicleros*) in Mexico and Central America, for example, develop a characteristic ulcerated lesion on their ear following infection by *L mexicana*.<sup>16</sup> CL lesions can be single or multiple and infection can spread through the lymphatic system, causing lymph node enlargement, satellite lesions, and even sporotrichoid lesions.<sup>17</sup> Atypical forms, such as eczematous, erysipeloid, lupoid, annular, and verrucous lesions, are more common in the New World.<sup>18</sup> CL lesions can resolve spontaneously within several months leaving a scar. Some cases, however, become chronic or spread. Chronic recurrent forms are typical in *L tropica* infections and are characterized by the formation of papules at the periphery of previously healed ulcers.<sup>19</sup> Chronicity has been linked to numerous factors, including increased arginase activity in polymorphonuclear leukocytes.<sup>20</sup> Diffuse CL, which is caused by *L aethiopica*, *L Mexicana*, or *L amazonensis*, presents as multiple nonulcerated papules and/or nodules involving most of the skin.<sup>5</sup> The lesions contain abundant parasites and can result in significant facial changes, causing a leonine-like appearance similar to that seen in lepromatous leprosy. Mucosal lesions are common.<sup>21</sup>

**Table 1** Clinical and Epidemiological Characteristics of the Main Species of *Leishmania*.

Complex	Main Species	Main Reservoir	Main Geographic Distribution	Most Common Clinical Patterns and Their Main Characteristics	Natural Progression
<i>Leishmania donovani</i>	<i>L. donovani</i>	Human	India, Bangladesh, Ethiopia, Sudan	VL: fever, hepatosplenomegaly, weight loss, anemia PKDL: macular, papular, or nodular lesions	Without treatment, death within 2 y  Resolves spontaneously in up to 85% of cases in Africa, but rarely in Asia
	<i>Leishmania infantum</i>	Dog, hare	Mediterranean Basin, China	CL: mildly inflammatory solitary nodules VL: more common in children and immunosuppressed individuals	Tends to resolve spontaneously within a year, conferring immunity
<i>Leishmania tropica</i>	<i>Leishmania infantum chagasi</i>	Dog, fox	Central and South America	CL: <3 dry ulcers, mostly on the head; chronic, recurrent course	Most cases resolve spontaneously within 2 y
	<i>L. tropica</i>	Human	Eastern Mediterranean, Middle East, India	LC: multiple moist inflammatory ulcers, most of which progress rapidly	70% of cases resolve in 4 mo but leave severe scarring
	<i>Leishmania major</i>	Rodents	Africa, Middle East, Central Asia, India, China	LC: localized nodules or ulcers; may result in diffuse involvement	Usually resolves spontaneously with 2-5 years, except for diffuse forms
<i>Leishmania mexicana</i>	<i>Leishmania aethiopica</i>	Hyraxes (dassie)	Ethiopia, Kenya	LC: solitary or multiple ulcerated lesions, sometimes with diffuse involvement; responsible for characteristic ulcerated ulcer lesion in gum collectors	>80% of cases resolve spontaneously within 3–4 mo, although some lesions can last up to 20 y
	<i>L. mexicana</i>	Rodents, marsupials	Mexico, Central and South America, Texas	LC: ulcerated lesions, often with diffuse involvement Can progress to MCL	Not well described
<i>Leishmania (Viannia) braziliensis</i>	<i>Leishmania amazonensis</i>	Rodents, possums	South America	LC: ulcerated lesions MCL: mainly caused by <i>L. braziliensis</i> ; ulcerated lesions in oral and nasal mucosa; can extend to oropharynx and larynx	Causes serious infections; often associated with satellite lesions, subcutaneous nodules, and enlarged locoregional lymph nodes; 6% of cases resolve spontaneously in 6 mo
	<i>Leishmania panamensis</i>	Sloths	Panama, Costa Rica, Colombia	CL: superficial ulcers, often with lymphatic spread MCL: nasopharyngeal involvement	Does not resolve spontaneously. Spreads through lymphatic vessels and can cause mucocutaneous lesions

Table 1 (Continued)

Complex	Main Species	Main Reservoir	Main Geographic Distribution	Most Common Clinical Patterns and Their Main Characteristics	Natural Progression
	<i>Leishmania guyanensis</i>	Possums, sloths, and anteaters	South America	LC: multiple ulcers that can spread through the lymphatic system, resulting in a sporotrichoid distribution MCL: cutaneous involvement that can progress to mucocutaneous involvement	Tends to need treatment and recurs frequently; at times, resolves spontaneously within 6 months

Abbreviations: CL, cutaneous leishmaniasis; MCL, mucocutaneous leishmaniasis; PKDL, post-kala-azar dermal leishmaniasis; VL, visceral leishmaniasis.

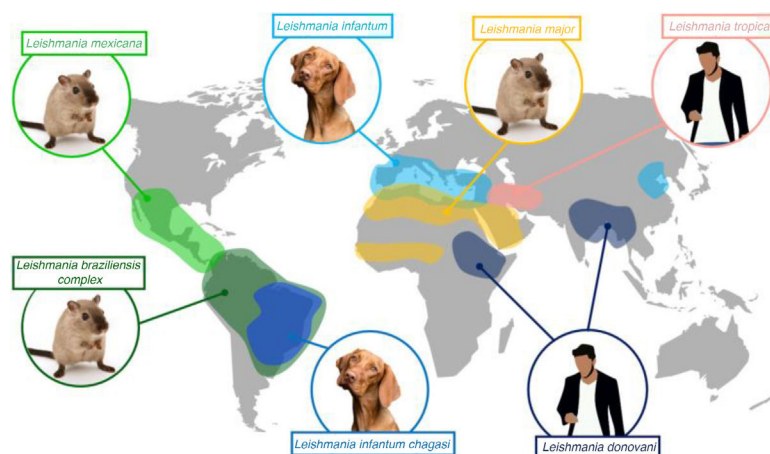


Fig. 1 Geographic distribution of the main *Leishmania* species and their principal reservoirs.

### Mucocutaneous Leishmaniasis

Mucosal involvement can coincide with cutaneous involvement or occur after the clearance of cutaneous lesions, sometimes even year later. The infection can spread through the bloodstream or lymphatic system. In endemic areas, mucosal involvement may be seen in up to 20% of patients.<sup>22</sup> Most cases of MCL are caused by *L. braziliensis*, but other species involved are *L. amazonensis*, *L. guyanensis*, and *L. panamensis*. The nasal (Fig. 3A) and oral mucosa are the most frequently affected areas. Lesions in the oral cavity can spread to the oropharynx and larynx, and may affect cartilage and vocal cords.<sup>23</sup> MCL lesions are ulcerated and can cause disfigurement. Treatment is essential to control infection, as the condition can be fatal.

### Visceral Leishmaniasis

In VL, infected macrophages spread through the reticuloendothelial system to the bone marrow, spleen, and liver. Clinical manifestations include fever, weight loss, hepatosplenomegaly, and lymph node enlargement. VL is mainly

caused by *L. donovani* in adults and *L. infantum* or *L. chagasi* in children and immunosuppressed individuals. Patients may also develop skin manifestations, which can be specific, such as papules (Fig. 3b), nodules, and ulcers, or nonspecific, such as purpura and hyperpigmentation (Fig. 3C). The presence of hyperpigmentation probably explains the origin of the term *kala-azar*, which means *black fever* in Hindu. Skin darkening has been described in 9.88% of patients with VL<sup>24</sup> and was recently linked to an increased production of cortisol.<sup>25</sup> Because hyperpigmented areas show amastigotes on histology, they could perhaps be considered a specific manifestation of leishmaniasis.<sup>26</sup>

### Post Kala-Azar Dermal Leishmaniasis

Post kala-azar dermal leishmaniasis (PKDL) occurs in patients with VL caused by *L. donovani* or, on occasions, *L. infantum*. It is particularly common in immunosuppressed patients<sup>27</sup> and can appear up to 20 years after treatment.<sup>28</sup> In patients infected with HIV, however, PKDL can coincide with, or at times precede, VL.<sup>29</sup> It is characterized by hypopigmented macules (Fig. 3D), flesh-colored nod-



**Fig. 2** A–C, The presence of an erythematous papule or nodule exhibiting progressive growth and a tendency to ulcerate in exposed areas, such as the face or extremities, is the most characteristic presentation of cutaneous leishmaniasis. D, Some patients have multiple lesions, or atypical presentations such as the verrucous lesion in this patient with New World cutaneous leishmaniasis. Clinical (E) and dermoscopic (F) image of a cutaneous leishmaniasis lesion on the forearm showing central ulceration surrounded by an erythematous area with peripheral polymorphous and hairpin vessels (asterisks) and white-yellow teardrop-like structures (arrows)(polarized light, original magnification  $\times 10$ ).

Images courtesy of Dr. Morales Moya (2C), Dr. Galimberti (2D), and Dr Mayo Martínez (2E and F).

ules and/or verrucous papules that mainly affect the face, although they can spread to the rest of the body.<sup>30</sup> PKDL mainly occurs in east Africa and India, although cases are occasionally seen in Spain.<sup>31</sup> The clinical presentation varies depending on geographic location and the host's immune response. In Asia, 90% of cases appear as macules, while in Africa, papules are predominant.<sup>30</sup> PKDL is more common—and severe—in immunosuppressed patients, who may exhibit atypical forms such as nodules not necessarily involving the face and a higher abundance of parasites in lesions.<sup>32</sup> The main entity in the differential diagnosis is leprosy, which is characterized by a loss of sensation. PKDL must be treated systemically, although African forms can resolve spontaneously within a year.

### HIV Coinfection

HIV infection has been linked to a 2000-fold increased risk of VL.<sup>33</sup> This is because both infections share a immunopathogenic mechanism involving macrophages and dendritic cells that accelerates progression in both cases.<sup>34</sup> HIV was responsible for the resurgence of VL in Europe during the 1990s that hit Spain, Portugal, Italy, and France particularly hard.<sup>33,35</sup> Eighty percent of all cases of VL-HIV coinfection reported to the WHO during this period were from Spain.<sup>36</sup> In addition, a retrospective analysis of all patients hospitalized for leishmaniasis in Spain between 1997 and 2008 found that 37% were HIV positive.<sup>37</sup> Coinfection can result in atypical presentations of CL and VL,<sup>18,38,39</sup>



**Fig. 3** A, Mucocutaneous leishmaniasis manifests as a chronic ulcer often affecting the nasal mucosa. B, Firm cutaneous leishmaniasis papules on the palm of a patient coinfecting with visceral leishmaniasis and human immunodeficiency virus. C, Hyperpigmented macules in the neck area. D, Hypopigmented macules as a manifestation of post-kala-azar dermal leishmaniasis in a 7-year-old boy. The diagnosis was established by molecular techniques, as the biopsy was negative.

poor response to treatment, higher mortality rates, higher viral loads, and faster progression to AIDS. The introduction of highly active antiretroviral therapy in 1997 increased survival rates and also reduced the incidence and recurrence of leishmaniasis.<sup>33</sup> It is important thus to rule out HIV in all patients with VL, even those living in endemic areas.<sup>3</sup>

### Leishmaniasis and TNF Inhibitors

TNF inhibitors are widely used to treat inflammatory diseases such as psoriasis and rheumatoid arthritis. TNF- $\alpha$  participates in cell-mediated immune response by activating CD4<sup>+</sup> and CD8<sup>+</sup> T cells. Together with other cytokines such as interleukin 12 and IFN- $\gamma$ , it has a key role in early infection control. Parasite DNA has been detected in the blood of up to 58% of healthy individuals in the Mediterranean, an endemic area with high levels of exposure to *Leishmania*, but the actual numbers of people who develop the disease are low.<sup>40</sup> It is therefore believed that most immunocompetent individuals are able to control infection before it manifests. Likewise, it has been suggested that TNF- $\alpha$  inhibitors could favor the reactivation of latent leishmaniasis,<sup>10</sup> and also modify clinical presentation, the natural course of disease, and response to treatment. Based on the cases published to date, the best approach to dealing with reactivation of a latent infection would appear to be to treat the infection with systemic therapy and interrupt TNF inhibitor treatment until the infection has cleared<sup>10</sup>; the treatment should then be reintroduced with close follow-up. Etanercept and

certolizumab appear to be associated with a lower risk of reactivation than adalimumab or infliximab.<sup>41</sup>

### Diagnosis

Leishmaniasis is diagnosed by demonstrating the presence of *Leishmania* amastigotes in clinical specimens using direct microscopic examination or molecular analysis based on nuclear or kinetoplast DNA amplification. Amastigotes are round and have a diameter of 1 to 4  $\mu\text{m}$  and a characteristic rod-shaped structure known as a *kinetoplast*. The main sampling and diagnostic techniques are summarized in Table 2.<sup>42-45</sup> Considering the limited sensitivity of some of the diagnostic techniques, it may be necessary to take several samples and/or use a combination of techniques to reach a diagnosis. In the case of biopsy specimens, for example, one part could be used for histology, another for touch imprint cytology, and another for culture. Diagnostic sensitivity in VL varies according to tissue type and can be as high as 90% for spleen tissue. Spleen aspiration is the procedure of choice for diagnosing VL, but it is associated with a high risk of intra-abdominal bleeding. Sensitivity rates ranging from 50% to 85% have been reported for bone marrow samples, and even lower rates have been described for lymph node and peripheral blood.<sup>46</sup> In PKDL, diagnostic sensitivity using smears or biopsy specimens is largely dependent on the type of lesion analyzed, with rates of up to 100% for nodular lesions contrasting with very low rates for macular lesions,<sup>47</sup> where more sensitive molecular methods are needed.<sup>48</sup> More information on sampling methods and diagnostic techniques

**Table 2** Sampling Methods and Diagnostic Techniques for Cutaneous and Mucocutaneous Leishmaniasis.

Type of Lesion	Sampling Method	Diagnostic Techniques	Technique	Considerations	
Ulcer	Base	Swab. To collect DNA. Rub ulcer several times.	PCR	Real-time PCR Samples can be stored in high concentrations of ethanol or in paraffin, although paraffin-embedded samples yield lower sensitivity	<i>Advantages:</i> highest sensitivity and specificity, relatively fast, and allows identification of species <i>Disadvantages:</i> not always available <i>Sensitivity:</i> 100%
		<i>Touch print cytology.</i> Remove the crust and take an imprint of the base of the ulcer on a slide.	Smear	Smear Giemsa staining and direct visualization using ×100 lens and oil immersion Visualization of amastigotes and their characteristic kinetoplast structure	<i>Advantages:</i> fast and cheap <i>Disadvantages:</i> does not allow identification of species <i>Sensitivity:</i> 85%
	Border	<i>Fine-needle aspiration.</i> Aspirate using a 1-mL syringe with a 20-25-G needle and 0.1 mL of sterile saline solution 0.9%. If no material is obtained, inject 0.05-0.1 mL of the saline solution and reaspirate.  <i>Skin scrapings.</i> Use a scalpel to obtain tissue scrapings from the upper dermis, via an incision or removal of necrotic tissue. Requires local anesthesia and bleeding control for better results. <i>Biopsy.</i> Shave or punch.	Smear, culture, PCR  Smear, culture, PCR	Culture Sterile sample Medium: Novy-MacNeal-Nicolle  Histology Hematoxylin-eosin	<i>Advantages:</i> allows identification of species and definitive diagnosis <i>Disadvantages:</i> frequent contamination by skin flora and low sensitivity <i>Sensitivity:</i> 40% Recent microculture techniques offer higher sensitivity (close to 100%), but do not allow identification of species  <i>Advantages:</i> can be used to rule out other possible causes, in particular, malignancy <i>Disadvantages:</i> invasive, does not allow identification of species, and has low sensitivity <i>Sensitivity:</i> 60%
Nodule/plaque	Fine-needle aspiration Biopsy	Smear, culture, PCR Smear, culture, PCR, histology			

Abbreviation: PCR: polymerase chain reaction.

can be consulted in the guidelines of the Centers for Disease Control and Prevention<sup>49</sup> and the Infectious Diseases Society of America.<sup>50</sup>

Histology can show nonspecific features such as ulceration, pseudoepitheliomatous hyperplasia, and a mixed inflammatory infiltrate, as well as specific features, such as amastigotes in dermal macrophages, seen in 50% to 70% of cases<sup>43</sup> (Fig. 4). As lesions develop, there is an increase in the number of giant cells and a decrease in that of parasites. Other findings include tuberculoid granulomas,<sup>51</sup> and in advanced stages, dermal fibrosis and abundant plasma cells.<sup>52</sup> Four histologic patterns have been described for leishmaniasis: 1) abundant amastigotes (45%); 2) a mixture of macrophages, neutrophils, and plasma cells accompanied by necrosis (27.5%); 3) incipient granulomas with epithelioid cells, lymphocytes, and plasma cells (15%); and 4) fully formed epithelioid granulomas with Langhans-type giant cells.<sup>53</sup> Histologic findings are similar in CL and MCL. A diffuse infiltrate of macrophages containing large numbers of amastigotes is seen in diffuse CL.<sup>52</sup>

Dermoscopy is another useful diagnostic aid.<sup>54</sup> The most frequently described dermoscopic structures are erythema (100%); vascular structures (90.6%), including polymorphous (40.2%), hairpin (39.4%), and arborizing vessels (38.6%); crusts (70.1%); and erosion/ulceration (44.1%). Less common but more characteristic structures are white-yellow teardrop-like structures (42.5%) and the white starburst pattern (8.6%)<sup>55</sup> (Fig. 2F).

Other tests, which are less useful in the diagnosis of CL, are the Montenegro skin test and serological tests. The former consists of the intradermal injection of leishmanin, and its results are read and interpreted in a similar way to those of the tuberculin test. It is negative in diffuse CL, active VL, and PKDL,<sup>56</sup> and cannot differentiate between current and past infection. It is mainly useful for epidemiological purposes.

Serological tests include the direct agglutination test, immunofluorescence, enzyme-linked immunoassay (ELISA), and Western blot analysis. These tests are highly sensitive in VL.<sup>57</sup> Antibody titers, however, drop very slowly after cure and the test does not discriminate between active and past infection. Results can also be affected by cross-reactivity with other antibodies (e.g., Chagas disease).<sup>58</sup> In addition, asymptomatic individuals in endemic areas often have antibodies, meaning that results have to be carefully interpreted according to the clinical context. High levels of anti- $\alpha$ -galactosyl antibodies have been detected using ELISA in CL caused by *L. tropica* and *L. major*.<sup>59</sup> A number of rapid diagnostic tests, such as rK39, are also available. These have high sensitivity for VL,<sup>60</sup> but they have the same limitations as other serological tests. More recent antigen detection tests include the latex agglutination test<sup>61</sup> and ELISA<sup>62</sup> with urine samples in VL and an immunochromatographic strip that tests for peroxidoxin antigen<sup>63</sup> in CL.

## Differential Diagnosis

The main entities that should be considered in the differential diagnosis are other infections, such as ecthyma, sporotrichosis, skin tuberculosis, furuncular myiasis, subcutaneous mycosis, tertiary syphilis, and

lepromatous leprosy; malignancies, such as squamous cell carcinoma, basal cell carcinoma, and lymphoma; and other skin disorders, such as persistent arthropod bite reactions, sarcoidosis, and granulomatosis with polyangitis.<sup>43</sup>

## Treatment

Many cases of CL resolve spontaneously in under 2 years. Variations in resolution times are largely related to the species involved (Table 1), with infections caused by *L. braziliensis* and *L. panamensis* most likely to persist.<sup>64</sup> Depending on this and other factors, such as anatomic location, infection severity, and the host's immune response, CL can be classified as simple or complex. Simple infections can be treated conservatively or with local treatments, while complex infections require systemic therapies.<sup>50</sup> The characteristics of simple and complex CL are summarized in Table 3. Multiple treatments exist for leishmaniasis, although the evidence supporting the options for CL is weak.<sup>65</sup> The main treatments are shown in Table 4.<sup>66-97</sup>

After weighing up the risks and benefits, a watch-and-wait approach can be taken in patients who meet the criteria for simple CL. Local treatment can be used for lesions that do not resolve spontaneously or when the goal is to achieve a faster cure or reduce the risk of scarring. The most widely accepted local treatments are intralésional pentavalent antimonials<sup>66-69</sup> and cryotherapy.<sup>76</sup> The combination of both produces better outcomes.<sup>70,71</sup> Topical paromomycin is also used to treat CL, particularly in the New World.<sup>73-75</sup>

There have been reports of good responses to other local treatments,<sup>98</sup> such as carbon dioxide laser therapy,<sup>99</sup> which has shown efficacy rates of 93% and very few adverse effects (hyperpigmentation, persistent erythema, and hypertrophic scarring); photodynamic therapy<sup>80</sup>; and imiquimod.<sup>100</sup>

Systemic therapies are recommended for patients who meet any of the criteria for complex CL. Liposomal amphotericin B is highly effective but carries a risk of nephrotoxicity.<sup>89-93</sup> The traditional use of systemic pentavalent antimonials<sup>85-88</sup> to treat complex CL and MCL has led to resistance in certain areas, limiting their use.<sup>87</sup> Other options include azoles,<sup>81</sup> miltefosine,<sup>82-84</sup> and pentamidine.<sup>94-96</sup>

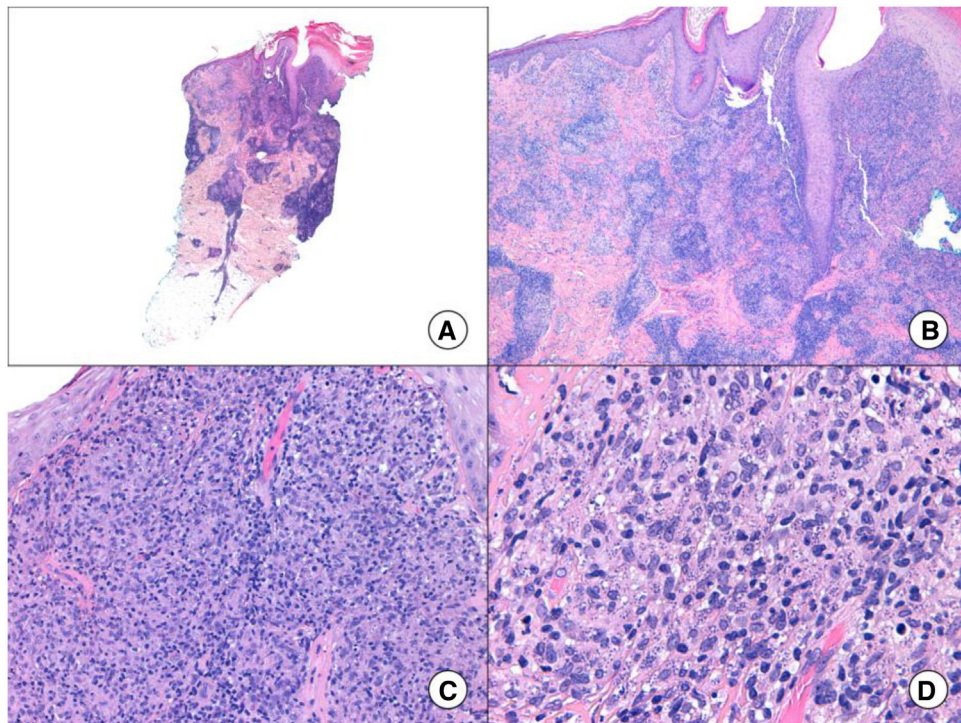
A clinical trial comparing twice-daily chloroquine 250 mg and once-daily doxycycline 200 mg for 3 months reported an efficacy of 100% and 92%, respectively.<sup>101</sup>

Annual check-ups are recommended in patients with CL due to *L. braziliensis* to enable early detection of progression to MCL. Signs of MCL include persistent nasal secretion or bleeding.

PKDL from Africa is not normally treated as the vast majority of cases (85%) resolve spontaneously within a year. In India, however, systemic treatment with miltefosine or amphotericin B is generally needed.<sup>1</sup>

We have drawn up an algorithm for the management of CL and MCL based on current guidelines and expert recommendations (Fig. 5).





**Fig. 4** A, Panoramic punch biopsy of an erythematous nodule on the forearm (hematoxylin-eosin, original magnification  $\times 5$ ). B, Dense superficial dermal inflammatory infiltrate and pseudoepitheliomatous hyperplasia (hematoxylin-eosin, original magnification  $\times 10$ ). C, Detail of inflammatory infiltrate composed mainly of macrophages, lymphocytes, and some epithelioid cells (hematoxylin-eosin, original magnification  $\times 20$ ). D, Characteristic amastigotes in infected macrophages (hematoxylin-eosin, original magnification  $\times 40$ ).

**Table 3** Characteristics of Simple and Complex Cutaneous Leishmaniasis.

Simple Cutaneous Leishmaniasis	Complex Cutaneous Leishmaniasis
<i>Leishmania</i> species associated with low likelihood of mucosal involvement	<i>Leishmania</i> species associated with high likelihood of mucosal involvement, particularly species from <i>Leishmaniasis braziliensis</i> complex
No notable mucosal involvement	Mucosal involvement, subcutaneous nodules, and/or significantly enlarged regional lymph nodes
Single or few lesions < 1 cm	$\geq 5$ lesions measuring > 1 cm or single lesion measuring > 5 cm
Located in areas without significant cosmetic concerns and that are accessible for local treatment	Located on the face, ears, fingers or toes, and skin covering joints or genitals
Immunocompetent host	Immunosuppressed host (HIV, TNF inhibitors, etc.)
Lesions showing signs of spontaneous resolution at diagnosis	History of non-response to local treatment
No criteria for complex cutaneous leishmaniasis	Recurrent disease or diffuse presentation

Abbreviations: HIV, human immunodeficiency virus; TNF, tumor necrosis factor.

## Prevention and Control

Prophylactic vaccines for leishmaniasis in humans are not yet available. The fact that most patients who recover from leishmaniasis do not become reinfected facilitates vaccine research efforts.<sup>102</sup> One of the main control strategies for CL and VL, apart from vector control, involves early detection and treatment, as infected patients are reservoirs. The WHO's target for CL in the eastern Mediterranean region was to detect 70% of all cases and to treat at least 90% by 2020, an ambitious target considering the few treatment options available, the suboptimal diagnostic tools, and the

low levels of awareness among the scientific community, particularly in relation to CL, which is not classified as an infection control priority.

## Conclusions

CL has become a relatively common condition in our setting due to international travel and migration. Its management, however, can be complicated due to low indices of suspicion among clinicians, the low sensitivity of some diagnostic tests, poor access to molecular testing, limited treatment options, and adverse treatment effects.

**Table 4** Main Treatments for Cutaneous and Mucocutaneous Leishmaniasis.

Treatment	Drug or Device	Indication	Dose, Adverse Effects and Other Considerations	Effectiveness <sup>97</sup>	Level of Evidence <sup>1, a</sup>
<b>Local Treatment</b>					
<i>Intralesional</i>					
Pentavalent antimonials	Sodium stibogluconate <sup>66-68</sup> (Pentostam 100 mg/mL, foreign medication)	OWCL Consider for NWCL if no risk of mucosal involvement	Intradermal injection 0.2-0.5 mL (0.1 mL/cm <sup>2</sup> in up to 5 sites/lesion every 3-7 d for up to 5 sessions) May require local anesthesia More effective when combined with cryotherapy <sup>70,71</sup> Intralesional administration does not usually cause adverse effects (except for local reactions), although QT interval prolongation has been reported in up to 25% of patients <sup>72</sup>	41%-98% (OWCL) 77%-90% (NWCL) 89%-91% (when combined with cryotherapy)	A (combined with cryotherapy, LCVM) B (NMCL)
	Meglumine antimoniate <sup>69</sup> (Glucantime 1500 mg/5 mL)				
<i>Topical</i> Paromomycin <sup>73</sup>	Paromomycin ointment <sup>74</sup> (Leshcutan, paromomycin 15% + MBCL 12%, foreign medication)	Ulcerated lesions due to <i>Leishmania</i> spp. in OWCL and NWCL	Twice daily for 10-20 d	Similar to intralesional pentavalent antimonials in OWCL, and inferior in NWCL	A (OWCL)
	Paromomycin 15% cream + gentamicin 0.5% <sup>75</sup> (not commercially available)	Better responses in <i>Leishmania major</i> and <i>Leishmania panamensis</i>			B (NWCL)
Cryotherapy <sup>76</sup>	Liquid nitrogen	Recent-onset CL lesions Residual lesions after systemic treatment	3 freeze-thaw cycles for 15-20 s, with halo of 1-2 mm; repeated every 3 wk until cure	57%-75% (similar efficacy to intralesional pentavalent antimonials)	A (combined with IL APV, OWCL)
		Consider in pregnant and breastfeeding women and patients with contraindications for systemic treatment	More effective when combined with intralesional pentavalent antimonials Can cause permanent hypopigmentation	89%-91% (if combined with intralesional pentavalent antimonials)	

Table 4 (Continued)

Treatment	Drug or Device	Indication	Dose, Adverse Effects and Other Considerations	Effectiveness <sup>97</sup>	Level of Evidence <sup>1, a</sup>
Thermotherapy <sup>77-79</sup>	Application of superficial heat via radiofrequency (ThermoMed)	OWCL and NWCL Consider in pregnant and breastfeeding women and patients with contraindications for systemic treatment	50 °C for 30 s applied to lesion and up to 1-2 mm of surrounding skin; 1-2 sessions Based on in vitro studies showing that <i>Leishmania</i> parasites do not multiply at > 39 °C Requires local anesthesia Produces second-degree burns	48%-98% (OWCL) 58%-90% (NWCL)	A (OWCL, NWCL)
PDT <sup>80</sup>	Conventional PDT with ALA or MAL	OWCL, especially that caused by <i>L major</i> or <i>Leishmania tropica</i> Consider in CL resistant to other treatments or in aesthetically sensitive areas Not recommended in CL due to <i>Leishmania braziliensis</i> complex or <i>Leishmania donovani</i> complex	1-7 sessions a week; ≥ 3 sessions more effective than ≤2 Mechanism of action believed to involve immune system response as does not directly kill the parasite Conventional PDT with ALA (6 sessions) seems to be at least as effective as cryotherapy (5 sessions) and produces better aesthetic results, although is more poorly tolerated Local adverse effects (burning, reddening, edema, pain)	96%-100% (conventional PDT) 74%-82% (daylight PDT)	B (conventional PDT, OWCL)
<b>Systemic therapy</b> <i>Oral</i>	Daylight PDT Intralesional PDT				
[1,0]Azoles <sup>81</sup>	Fluconazole	Off-label use for CL with lymphatic involvement and certain complex cases of CL; not indicated for cases due to <i>L braziliensis</i>	200-400 mg/d for 42 d Adverse effects include gastrointestinal disturbances and hepatotoxicity	44%-81% (OWCL) 22% (NWCL)	A (OWCL, fluconazole 200 mg/d, 42 d, <i>Lmajor</i> )
Miltefosine <sup>82-84</sup>	Itraconazole Miltefosine (Impavido 50 mg capsules, foreign medication)	LC due to <i>L braziliensis</i> complex; can also be used in LC due to <i>L major</i> , <i>L tropica</i> , or <i>Leishmania mexicana</i>	200 mg/d for 42-56 d 2.5 mg/kg/d (maximum 150 mg), administered orally in 3 doses for 28 d Expensive Teratogenic and can cause gastrointestinal disturbances and hepatotoxicity	81% (OWCL) 53%-91% (NWCL)	B (NWCL) C (OWCL)

(Continued)

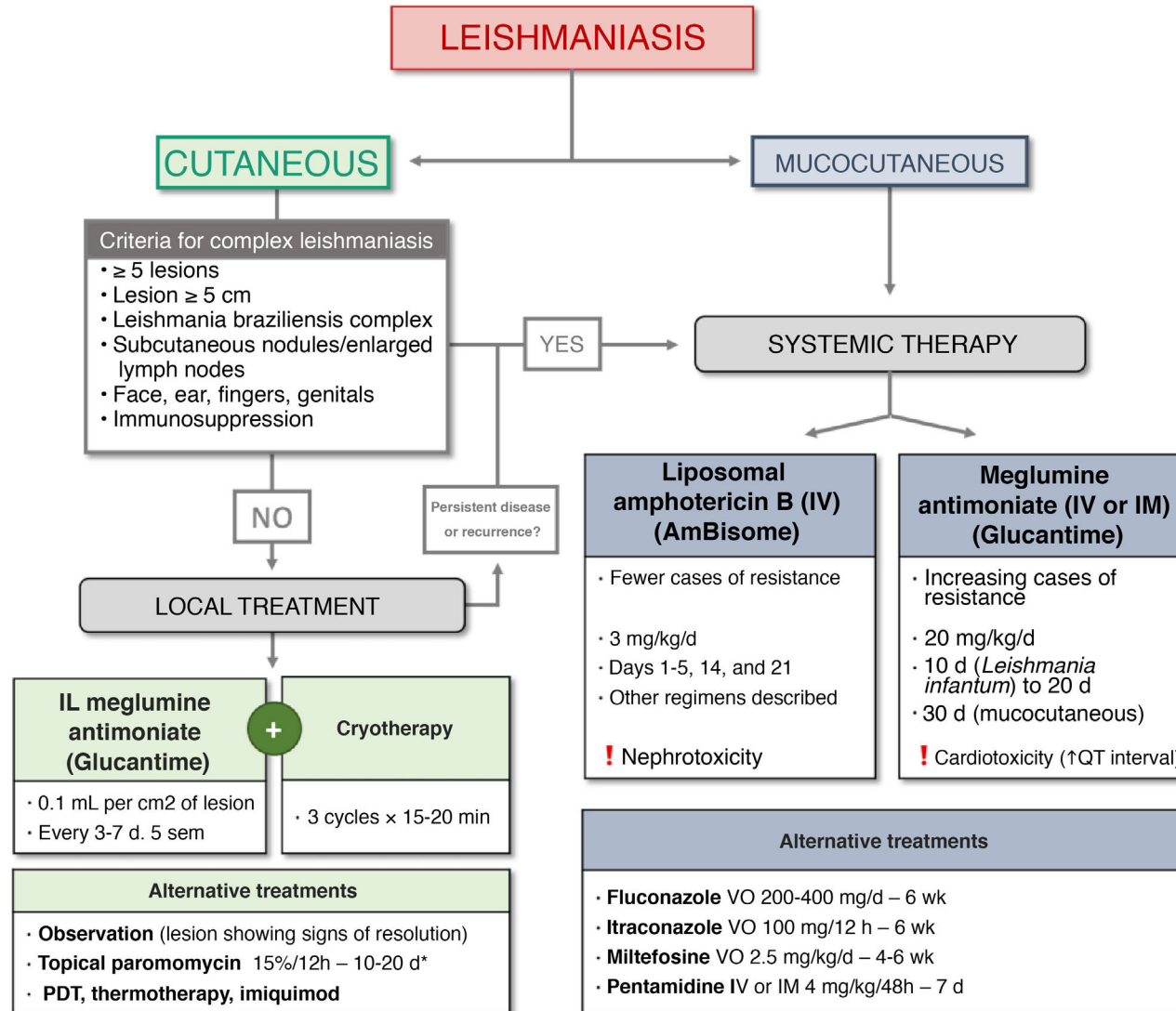
Treatment	Drug or Device	Indication	Dose, Adverse Effects and Other Considerations	Effectiveness <sup>97</sup>	Level of Evidence <sup>1,a</sup>
<i>Parenteral</i> Pentavalent antimonials <sup>85-88</sup>	Sodium stibogluconate (Pentostam, 100 mg/mL)	[1,0]Complex OWCL and NWCL and MCL	20 mg/kg/d in a single daily dose administered orally (preferred) or intramuscularly (very painful) for 10-30 d Treatment failures have increased due to resistance <sup>87</sup> Usually associated with pentoxifylline 400 mg 3 times a day for 10-20 d, or allopurinol 20 mg/kg for 30 days in recurrent LC due to <i>L tropica</i> Can cause cardiotoxicity (e.g., arrhythmias, QT interval prolongation), pancreatitis, and hepatotoxicity	41%-85% (OWCL) 77%-90% (NWCL) 30%-90% (MCL)	A (OWCM, MCL) C (recurrent CL)
Amphotericin B <sup>89-93</sup>	Meglumine antimoniate (Glucantime 1500 mg/5 mL) Liposomal (AmBisome 50 mg), treatment of choice due to its lower toxicity compared with conventional options	Off-label use for complex CL and MCL	3 mg/kg/d IV for 7 d (days 1-5, 14, and 21) up to a total dose of 21 mg/kg. Optimal dose for LC not well-defined; treatment regimens are based on the doses used for VL; higher doses and longer treatments are usually required in immunosuppressed patients Approval of topical formulation of amphotericin 3% with similar efficacy to intralesional pentavalent antimonials in LC <sup>92</sup> Can cause nephrotoxicity, lower back pain, hypokalemia, chills	84%-100% (OWCL, NWCL) 78%-100% (MCL)	C (NWCL, MCL)

(Continued)

Treatment	Drug or Device	Indication	Dose, Adverse Effects and Other Considerations	Effectiveness <sup>97</sup>	Level of Evidence <sup>1,a</sup>
Pentamidine <sup>94-96</sup>	Pentamidine isethionate 300 mg (Pentacarinat)	Alternative treatment for NWCL; good responses in <i>Leishmania guyanensis</i>	4 mg/kg/d, intramuscular on alternating days for a week Can also be administered intravenously (more effective). Intralesional administration recently approved; similar efficacy to pentavalent antimonials Can cause severe toxicity: pancreatitis, QT interval prolongation, hyperpotassemia	58%-95% (NWCL) 91% (MCL, but 25% recurrence rate)	C (NWCL, MCL)

Abbreviations: ALA, aminolevulinic acid; IL, intralesional; MAL, methyl aminolevulinate; MBCL, methyl benzethonium chloride; MCL, mucocutaneous leishmaniasis; NWCL, New World cutaneous leishmaniasis; OWCL, Old World cutaneous leishmaniasis; PDT, photodynamic therapy; VL, visceral leishmaniasis.

<sup>a</sup> Level of evidence (World Health Organization system): evidence supported by A, at least 1 well-designed clinical trial; B, well-designed nonrandomized clinical trials; C, descriptive studies, expert committees; D, expert opinions without conclusive studies.



**Fig. 5** Treatment algorithm for cutaneous and mucocutaneous leishmaniasis. IL indicates intralésional; IM, intramuscular; IV, intravenosa; PDT, photodynamic therapy; VO, oral. \* foreign medication.

HIV coinfection and use of TNF inhibitors are associated with atypical presentations and the need for systemic treatment.

## Conflicts of Interest

The authors declare that they have no conflicts of interest.

## References

- World Health Organization [Accessed 20 October 2020]. Available from: <http://www.who.int/mediacentre/factsheets/fs375/es/>, 2020.
- Lupi O, Bartlett BL, Haugen RN, Dy LC, Sethi A, Klaus SN, et al. Tropical dermatology: Tropical diseases caused by protozoa. *J Am Acad Dermatol*. 2009;60:897–928.
- Burza S, Croft SL, Boelaert M. Leishmaniasis. *Lancet*. 2018;392:951–70.
- Kevric I, Cappel MA, Keeling JH. New World and Old World Leishmania Infections: A Practical Review. *Dermatol Clin*. 2015;33:579–93.
- Handler MZ, Patel PA, Kapila R, Al-Qubati Y, Schwartz RA. Cutaneous and mucocutaneous leishmaniasis: Clinical perspectives. *J Am Acad Dermatol*. 2015;73:897–908.
- Suárez Rodríguez B, Isidoro Fernández B, Santos Sanz S, Sierra Moros MJ, Molina Moreno R, Astray Mochales J, et al. Situación epidemiológica y de los factores de riesgo de transmisión de *Leishmania infantum* en España. *Rev Esp Salud Publica*. 2012;86:555–64.
- Gomez-Barroso D, Herrador Z, San Martín JV, Gherasim A, Aguado M, Romero-Mate A, et al. Spatial distribution and cluster analysis of a leishmaniasis outbreak in the south-western Madrid region, Spain, September 2009 to April 2013. *Euro Surveill*. 2015;20:11–20.
- Galán-Puchades MT, Gómez-Samblás M, Suárez-Morán JM, Osuna A, Sanxis-Furió J, Pascual J, et al. Leishmaniasis in Norway Rats in Sewers, Barcelona, Spain. *Emerg Infect Dis*. 2019;25:1222–4.
- Alvar J, Canavate C, Gutierrez-Solar B, Jiménez M, Laguna F, López-Vélez R, et al. Leishmania and human immunodeficiency virus coinfection: the first 10 years. *Clin Microbiol Rev*. 1997;10:298–319.
- Bosch-Nicolau P, Ubals M, Salvador F, Sánchez-Montalvá A, Aparicio G, Erra A, et al. Leishmaniasis and tumor necrosis factor alpha antagonists in the Mediterranean basin. A switch in clinical expression. *PLoS Negl Trop Dis*. 2019;13:e0007708.
- Boggild AK, Caumes E, Grobusch MP, Schwartz E, Hynes NA, Libman M, et al. GeoSentinel Surveillance Network. Cutaneous and mucocutaneous leishmaniasis in travellers and migrants: a 20-year GeoSentinel Surveillance Network analysis. *J Travel Med*. 2019;26:taz055.
- Reithinger R, Dujardin JC, Louzir H, Pirmez C, Alexander B, Brooker S. Cutaneous leishmaniasis. *Lancet Infect Dis*. 2007;7:581–96.
- Wirth DF, Rogers WO, Barker R Jr, Dourado H, Suesebang L, Albuquerque B. Leishmaniasis and malaria: new tools for epidemiologic analysis. *Science*. 1986;234:975–9.
- Colmenares M, Kar S, Goldsmith-Pestana K, McMahon-Pratt D. Mechanisms of pathogenesis: differences amongst Leishmania species. *Trans R Soc Trop Med Hyg*. 2002;96:3–7.
- Scott P, Novais FO. Cutaneous leishmaniasis: immune responses in protection and pathogenesis. *Nat Rev Immunol*. 2016;16:581–92.
- Quijano-Pitman F. Identificación de la Leishmania trópica mexicana como agente etiológico de la Úlcera de los Chicleros. *Gac Med Mex*. 1999;135:331.
- Carvalho LMV, Pimentel MIF, Conceição-Silva F, Vasconcellos ÉCFE, Valette-Rosalino CM, Lyra MR, et al. Sporotrichoid leishmaniasis: a cross-sectional clinical, epidemiological and laboratory study in Rio de Janeiro State, Brazil. *Rev Inst Med Trop Sao Paulo*. 2017;59:e33.
- Meireles CB, Maia LC, Soares GC, Teodoro IPP, Gadelha MDSV, da Silva CGL, et al. Atypical presentations of cutaneous leishmaniasis: A systematic review. *Acta Trop*. 2017;172:240–54.
- Gitari JW, Nzou SM, Wamunyokoli F, Kinyeru E, Fujii Y, Kaneko S, et al. Leishmaniasis recidivans by *Leishmania tropica* in Central Rift Valley Region in Kenya. *Int J Infect Dis*. 2018;74:109–16.
- Mortazavi H, Sadeghipour P, Taslimi Y, Habibzadeh S, Zali F, Zahedifard F, et al. Comparing acute and chronic human cutaneous leishmaniasis caused by *Leishmania major* and *Leishmania tropica* focusing on arginase activity. *J Eur Acad Dermatol Venereol*. 2016;30:2118–21.
- Mariz BALA, Sánchez-Romero C, Alvarado NAP, Campos EMM, Almeida OP, Martínez-Pedraza R, et al. Diffuse cutaneous leishmaniasis with oral involvement in a patient of Northern Mexico. *Trop Doct*. 2019;49:303–6.
- David C, Dimier-David L, Vargas F, Torrez M, Dedet JP. Fifteen years of cutaneous and mucocutaneous leishmaniasis in Bolivia: a retrospective study. *Trans R Soc Trop Med Hyg*. 1993;87:7–9.
- Marra F, Chiappetta MC, Vincenti V. Ear, nose and throat manifestations of mucocutaneous Leishmaniasis: a literature review. *Acta Biomed*. 2014;85:3–7.
- Sarker CB, Chowdhury KS, Siddiqui NI, Jamal MF, Rahman S, Momen A, et al. Clinical profile of Kala-azar in adults: as seen in Mymensingh Medical College Hospital, Mymensingh, Bangladesh. *Mymensingh Med J*. 2003;12:41–4.
- Elkhair EB. Elevated cortisol level due to visceral leishmaniasis and skin hyper-pigmentation are causally related. *Int J Sci Commer Humanit*. 2014;2:86–92.
- Abadías-Granado I, Navarro-Bielsa A, Ferrando-Lamana L, et al. Hyperpigmentation as a guiding sign for the diagnosis of visceral leishmaniasis in a patient with human immunodeficiency virus (HIV). *Int J Dermatol*. 2021. Feb 1 <https://doi.org/10.1111/ijd.15429>
- Stark D, Pett S, Marriott D, Harkness J. Post-kala-azar dermal leishmaniasis due to *Leishmania infantum* in a human immunodeficiency virus type 1-infected patient. *J Clin Microbiol*. 2006;44:1178–80.
- Salotra P, Sreenivas G, Beena KR, Mukherjee A, Ramesh V. Parasite detection in patients with post kala-azar dermal leishmaniasis in India: a comparison between molecular and immunological methods. *J Clin Pathol*. 2003;56:840–3.
- Zijlstra EE. Biomarkers in Post-kala-azar Dermal Leishmaniasis. *Front Cell Infect Microbiol*. 2019;9:228.
- Zijlstra EE, Musa AM, Khalil EA, el-Hassan IM, el-Hassan AM. Post-kala-azar dermal leishmaniasis. *Lancet Infect Dis*. 2003;3:87–98.
- de Juan Martín F, Justa Roldán ML, Sáez de Adana Pérez E, Navarro Serrano E, Bouthelier Moreno M, Gilaberte Calzada Y, et al. Leishmaniasis dérmica postkala-azar [Postkala-azar dermal leishmaniasis]. *An Esp Pediatr*. 1997;46:63–4.
- Zijlstra EE. PKDL and other dermal lesions in HIV co-infected patients with leishmaniasis: review of clinical presentation in relation to immune responses. *PLoS Negl Trop Dis*. 2014;8:e3258.
- Alvar J, Aparicio P, Aseffa A, Den Boer M, Cañavate C, Dedet JP, et al. The relationship between leishmaniasis and AIDS: the second 10 years. *Clin Microbiol Rev*. 2008;21:334–59.

34. Mock DJ, Hollenbaugh JA, Daddacha W, Overstreet MG, Lazarski CA, Fowell DJ, et al. Leishmania induces survival, proliferation and elevated cellular dNTP levels in human monocytes promoting acceleration of HIV co-infection. *PLoS Pathog.* 2012;8:e1002635.
35. Monge-Maillo B, Norman FF, Cruz I, Alvar J, López-Vélez R. Visceral leishmaniasis and HIV coinfection in the Mediterranean region. *PLoS Negl Trop Dis.* 2014;8:e3021.
36. Desjeux P, Alvar J. Leishmania/HIV co-infections: epidemiology in Europe. *Ann Trop Med Parasitol.* 2003;97 Suppl 1:3–15.
37. Gil-Prieto R, Walter S, Alvar J, de Miguel AG. Epidemiology of leishmaniasis in Spain based on hospitalization records (1997-2008). *Am J Trop Med Hyg.* 2011;85:820–5.
38. Ejara ED, Lynen L, Boelaert M, Van Griensven J. Challenges in HIV and visceral leishmania co-infection: future research directions. *Trop Med Int Health.* 2010;15:1266–7.
39. Guiguemde RT, Sawadogo OS, Bories C, et al. *Leishmania major* and HIV co-infection in Burkina Faso. *Trans R Soc Trop Med Hyg.* 2003;97:168–9.
40. Mary C, Faraut F, Drogoul MP, Xeridat B, Schleinitz N, Cuisenier B, et al. Reference values for *Leishmania infantum* parasitemia in different clinical presentations: quantitative polymerase chain reaction for therapeutic monitoring and patient follow-up. *Am J Trop Med Hyg.* 2006;75:858–63.
41. Arens K, Filippis C, Kleinfelder H, Goetzee A, Reichmann G, Crauwels P, et al. Anti-Tumor Necrosis Factor  $\alpha$  Therapeutics Differentially Affect *Leishmania* infection of human macrophages. *Front Immunol.* 2018;9:1772.
42. Aronson NE, Joya CA. Cutaneous leishmaniasis: updates in diagnosis and management. *Infect Dis Clin North Am.* 2019;33:101–17.
43. Handler MZ, Patel PA, Kapila R, Al-Qubati Y, Schwartz RA. Cutaneous and mucocutaneous leishmaniasis: Differential diagnosis, diagnosis, histopathology, and management. *J Am Acad Dermatol.* 2015;73:911–28.
44. Elmahallawy EK, Sampedro Martinez A, Rodriguez-Granger J, Hoyos-Mallecot Y, Agil A, Navarro Mari JM, et al. Diagnosis of leishmaniasis. *J Infect Dev Ctries.* 2014;8:961–72.
45. Pagheh A, Fakhar M, Mesgarian F, Gholami S, Ahmadvour E. An improved microculture method for diagnosis of cutaneous leishmaniasis. *J Parasit Dis.* 2014;38:347–51.
46. Srivastava P, Dayama A, Mehrotra S, Sundar S. Diagnosis of visceral leishmaniasis. *Trans R Soc Trop Med Hyg.* 2011;105:1–6.
47. Salotra P, Singh R. Challenges in the diagnosis of post kala-azar dermal leishmaniasis. *Indian J Med Res.* 2006;123:295–310.
48. Adams ER, Versteeg I, Leeflang MM. Systematic review into diagnostics for post-kala-azar dermal leishmaniasis (PKDL). *J Trop Med.* 2013;2013:150746.
49. U. S. Centers for Disease Control and Prevention [Accessed 16 November 2020]. Available from: <https://www.cdc.gov/parasites/leishmaniasis/resources/pdf/cdc diagnosis guide leishmaniasis2016.pdf>, 2016.
50. Aronson N, Herwaldt BL, Libman M, Pearson R, Lopez-Velez R, Weina P, et al. Diagnosis and Treatment of Leishmaniasis: Clinical Practice Guidelines by the Infectious Diseases Society of America (IDSA) and the American Society of Tropical Medicine and Hygiene (ASTMH). *Am J Trop Med Hyg.* 2017;96:24–45.
51. Aoun J, Habib R, Charaffeddine K, Taraif S, Loya A, Khalifeh I. Caseating Granulomas in Cutaneous Leishmaniasis. *PLoS Negl Trop Dis.* 2014;8:e3255.
52. Mehregan DR, Mehregan DA, Mehregan AH. Histopathology of cutaneous leishmaniasis. *Gulf J Dermatol Venereol.* 1997;4:1–9.
53. Venkataram M, Moosa M, Devi L. Histopathological spectrum in cutaneous leishmaniasis: a study in Oman. *Indian J Dermatol Venereol Leprol.* 2001;67:294–8.
54. Llambrich A, Zaballos P, Terrasa F, Torne I, Puig S, Malvehy J, et al. Dermoscopy of cutaneous leishmaniasis. *Br J Dermatol.* 2009;160:756–61.
55. Ayhan E, Ucmak D, Baykara SN, Akkurt ZM, Arica M. Clinical and dermoscopic evaluation of cutaneous leishmaniasis. *Int J Dermatol.* 2015;54:193–201.
56. Momeni Boroujeni A, Aminjavaheri M, Moshtaghian B, Momeni A, Momeni AZ, et al. Reevaluating leishmanin skin test as a marker for immunity against cutaneous leishmaniasis. *Int J Dermatol.* 2013;52:827–30.
57. Cota GF, de Sousa MR, Demarqui FN, Rabello A. The diagnostic accuracy of serologic and molecular methods for detecting visceral leishmaniasis in HIV infected patients: meta-analysis. *PLoS Negl Trop Dis.* 2012;6:e1665.
58. Vexenat A, de C, Santana JM, Texeira AR. Cross-reactivity of antibodies in human infections by the kinetoplastid protozoa *Trypanosoma cruzi*, *Leishmania chagasi* and *Leishmania (vanni) braziliensis*. *Rev Inst Med São Paulo.* 1996;38:177–85.
59. Al-Salem WS, Ferreira DM, Dyer NA, Alyamani EJ, Balghonaim SM, Al-Mehna AY, et al. Detection of high levels of anti-alpha-galactosyl antibodies in sera of patients with Old World cutaneous leishmaniasis: a possible tool for diagnosis and biomarker for cure in an elimination setting. *Parasitology.* 2014;141:1898–903.
60. Boelaert M, Verdonck K, Menten J, Sunyoto T, van Griensven J, Chappuis F, et al. Rapid tests for the diagnosis of visceral leishmaniasis in patients with suspected disease. *Cochrane Database Syst Rev.* 2014;6. CD009135.
61. Ghatei MA, Hatam GR, Hossini MH, Sarkari B. Performance of latex agglutination test (KAtex) in diagnosis of visceral leishmaniasis in Iran. *Iran J Immunol.* 2009;6:202–7.
62. Ghosh P, Bhaskar KR, Hossain F, Khan MA, Vallur AC, Duthie MS, et al. Evaluation of diagnostic performance of rK28 ELISA using urine for diagnosis of visceral leishmaniasis. *Parasit Vectors.* 2016;9:383.
63. De Silva G, Somaratne V, Senaratne S, Vipuladasa M, Wickremasinghe R, Wickremasinghe R, et al. Efficacy of a new rapid diagnostic test kit to diagnose Sri Lankan cutaneous leishmaniasis caused by *Leishmania donovani*. *PLoS One.* 2017;12:e0187024.
64. Cota GF, de Sousa MR, Fereguetti TO, Saleme PS, Alvarisa TK, Rabello A. The cure rate after placebo or no therapy in American cutaneous leishmaniasis: a systematic review and meta-analysis. *PLoS One.* 2016;11:e0149697.
65. Gonzalez U, Pinart M, Reveiz L, Alvar J. Interventions for Old World cutaneous leishmaniasis. *Cochrane Database Syst Rev.* 2008;4:CD005067.
66. Ranawaka RR, Weerakoon HS, de Silva SHP. Randomized, double-blind, controlled, comparative study on intralesional 10% and 15% hypertonic saline versus intralesional sodium stibogluconate in *Leishmania donovani* cutaneous leishmaniasis. *Int J Dermatol.* 2015;54:555–63.
67. Tallab TM, Bahamdah KA, Mirdad S, Johargi H, Mourad MM, Ibrahim K. Cutaneous leishmaniasis: schedules for intralesional treatment with sodium stibogluconate. *Int J Dermatol.* 1996;35:594–7.
68. Bumb RA, Mehta RD, Ghiya BC, Jakhar R, Prasad N, Soni P, et al. Efficacy of short-duration (twice weekly) intralesional sodium stibogluconate in treatment of cutaneous Leishmaniasis in India. *Br J Dermatol.* 2010;163:854–8.
69. Brito NC, Rabello A, Cota GF. Efficacy of pentavalent antimoniate intralesional infiltration therapy for cutaneous leishmaniasis: A systematic review. *PLoS One.* 2017;12:e0184777.
70. Asilian A, Sadeghinia A, Faghihi G, Momeni A. Comparative study of the efficacy of combined cryotherapy and intrale-



- sional meglumine antimoniate (Glucantime) vs cryotherapy and intralesional meglumine antimoniate (Glucantime) alone for the treatment of cutaneous leishmaniasis. *Int J Dermatol.* 2004;43:281–3.
71. Salmanpour R, Razmavar MR, Abtahi N. Comparison of intralesional meglumine antimoniate, cryotherapy and their combination in the treatment of cutaneous leishmaniasis. *Int J Dermatol.* 2006;45:1115–6.
  72. Fernandes HJ, da Silva RE, Ramalho DB, Aguiar MG, Silveira JN, Cota G. Safety profile of meglumine antimoniate intralesional infiltration for cutaneous leishmaniasis. *Expert Rev Anti Infect Ther.* 2020;18:381–7.
  73. Kim DH, Chung HJ, Bleys J, Ghohestani RF. Is paromomycin an effective and safe treatment against cutaneous leishmaniasis? A meta-analysis of 14 randomized controlled trials. *PLoS Negl Trop Dis.* 2009;3:e381.
  74. Sosa N, Pascale JM, Jiménez AI, Norwood JA, Kreishman-Detrick M, Weina PJ, et al. Topical paromomycin for New World cutaneous leishmaniasis. *PLoS Negl Trop Dis.* 2019;13:e0007253.
  75. Ben Salah A, Ben Messaoud N, Guedri E, Zaatour A, BenAlaya N, Bettaieb J, et al. Topical paromomycin with or without gentamicin for cutaneous leishmaniasis. *N Engl J Med.* 2013;368:524–32.
  76. López-Carvajal L, Cardona-Arias JA, Zapata-Cardona MI, Sánchez-Giraldo V, Vélez ID, et al. Efficacy of cryotherapy for the treatment of cutaneous leishmaniasis: meta-analyses of clinical trials. *BMC Infect Dis.* 2016;16:360.
  77. Aronson NE, Wortmann GW, Byrne WR, Howard RS, Bernstein WB, Marovich MA, et al. A randomized controlled trial of local heat therapy versus intravenous sodium stibogluconate for the treatment of cutaneous *Leishmania major* infection. *PLoS Negl Trop Dis.* 2010;4:e628.
  78. David JR. The successful use of radiofrequency-induced heat therapy for cutaneous leishmaniasis: a review. *Parasitology.* 2018;145:527–36.
  79. Cardona-Arias JA, Vélez ID, López-Carvajal L. Efficacy of thermotherapy to treat cutaneous leishmaniasis: a meta-analysis of controlled clinical trials. *PLoS One.* 2015;10(5):e0122569.
  80. Morton CA, Szeimies R-M, Basset-Séguin N, Calzavara-Pinton PG, Gilaberte Y, Haedersdal M, et al. European Dermatology Forum guidelines on topical photodynamic therapy 2019 Part 2: emerging indications – field cancerization, photorejuvenation and inflammatory/infective dermatoses. *J Eur Acad Dermatol Venereol.* 2020;34:17–29.
  81. Galvao EL, Rabello A, Cota GF. Efficacy of azole therapy for tegumentary leishmaniasis: a systematic review and meta-analysis. *PLoS One.* 2017;12(10):e0186117.
  82. Mosimann V, Blazek C, Grob H, Chaney M, Neumayr A, Blum J. Miltefosine for mucosal and complicated cutaneous Old World leishmaniasis: a case series and review of the literature. *Open Forum Infect Dis.* 2016;3:ofw008.
  83. Sampaio RNR, Silva JSFE, de Paula CDR, Porto C, Motta JOCD, Pereira LIA, et al. A randomized, open-label clinical trial comparing the long-term effects of miltefosine and meglumine antimoniate for mucosal leishmaniasis. *Rev Soc Bras Med Trop.* 2019;52:e20180292.
  84. Iranpour S, Hosseinzadeh A, Alipour A. Efficacy of miltefosine compared with glucantime for the treatment of cutaneous leishmaniasis: a systematic review and meta-analysis. *Epidemiol Health.* 2019;41:e2019011.
  85. Firdous R, Yasinzai M, Ranja K. Efficacy of glucantime in the treatment of Old World cutaneous leishmaniasis. *Int J Dermatol.* 2009;48:758–62.
  86. Monge-Maillo B, López-Vélez R. Therapeutic options for old world cutaneous leishmaniasis and new world cutaneous and mucocutaneous leishmaniasis. *Drugs.* 2013;73:1889–920.
  87. Arevalo J, Ramirez L, Adai V, Zimic M, Tulliano G, Miranda-Verástegui C, et al. Influence of *Leishmania (Viannia)* species on the response to antimonial treatment in patients with American tegumentary leishmaniasis. *J Infect Dis.* 2007;195:1846–51.
  88. Cincurá C, de Lima CMF, Machado PRL, Oliveira-Filho J, Glesby MJ, Lessa MM, et al. Mucosal leishmaniasis: a retrospective study of 327 cases from an endemic area of *Leishmania (Viannia) braziliensis*. *Am J Trop Med Hyg.* 2017;97:761–6.
  89. Shirzadi MR. Liposomal amphotericin B: a review of its properties, function, and use for treatment of cutaneous leishmaniasis. *Res Rep Trop Med.* 2019;10:11–8.
  90. Guery R, Henry B, Martin-Blondel G, Rouzaud C, Cordoliani F, Harms G, et al. Liposomal amphotericin B in travelers with cutaneous and mucocutaneous leishmaniasis: not a panacea. *PLoS Negl Trop Dis.* 2017;11:e0006094.
  91. Wortmann G, Zapor M, Ressler R, Fraser S, Hartzell J, Pierson J, et al. Liposomal amphotericin B for treatment of cutaneous leishmaniasis. *Am J Trop Med Hyg.* 2010;83:1028–33.
  92. López L, Vélez I, Asela C, Cruz C, Alves F, Robledo S, et al. A phase II study to evaluate the safety and efficacy of topical 3% amphotericin B cream (Anfoleish) for the treatment of uncomplicated cutaneous leishmaniasis in Colombia. *PLoS Negl Trop Dis.* 2018;12:e0006653.
  93. Solomon M, Baum S, Barzilai A, Scope A, Trau H, Schwartz E, et al. Liposomal amphotericin B in comparison to sodium stibogluconate for cutaneous infection due to *Leishmania braziliensis*. *J Am Acad Dermatol.* 2007;56:612–6.
  94. Neves LO, Talhari AC, Gadelha EPN, Silva Júnior RM, Guerra JA, Ferreira LC, et al. A randomized clinical trial comparing meglumine antimoniate, pentamidine and amphotericin B for the treatment of cutaneous leishmaniasis by *Leishmania guyanensis*. *An Bras Dermatol.* 2011;86:1092–101.
  95. Christen J-R, Bourreau E, Demar M, Lightburn E, Couppié P, Ginouvés M, et al. Use of the intramuscular route to administer pentamidine isethionate in *Leishmania guyanensis* cutaneous leishmaniasis increases the risk of treatment failure. *Travel Med Infect Dis.* 2018;24:31–6.
  96. Soto J, Paz D, Rivero D, Soto P, Quispe J, Toledo J, et al. Intralesional pentamidine: A novel therapy for single lesions of Bolivian cutaneous leishmaniasis. *Am J Trop Med Hyg.* 2016;94:852–6.
  97. Chakravarty J, Sundar S. Current and emerging medications for the treatment of leishmaniasis. *Expert Opin Pharmacother.* 2019;20:1251–65.
  98. Nassif PW, De Mello TFP, Navasconi TR, Mota CA, Demarchi IG, Aristides SMA, et al. Safety and efficacy of current alternatives in the topical treatment of cutaneous leishmaniasis: a systematic review. *Parasitology.* 2017;144:995–1004.
  99. Shamsi Meymandi S, Zandi S, Aghaie H, Heshmatkhan A. Efficacy of CO(2) laser for treatment of anthroponotic cutaneous leishmaniasis, compared with combination of cryotherapy and intralesional meglumine antimoniate. *J Eur Acad Dermatol Venereol.* 2011;25:587–91.
  100. Firooz A, Khamesipour A, Ghoorchi MH, Nassiri-Kashani M, Eskandari SE, Khatami A, et al. Imiquimod in combination with meglumine antimoniate for cutaneous leishmaniasis: a randomized assessor-blind controlled trial. *Arch Dermatol.* 2006;142:1575–9.

101. Malik F, Hanif MM, Mustafa G. Comparing the Efficacy of Oral Chloroquine versus Oral Tetracycline in the Treatment of Cutaneous Leishmaniasis. *J Coll Physicians Surg Pak.* 2019;29:403–5.
102. Alvar J, Croft SL, Kaye P, Khamesipour A, Sundar S, Reed SG, et al. Case study for a vaccine against leishmaniasis. *Vaccine.* 2013;31 suppl 2:B244–9.